Effects of a Spirulina-Based Dietary Supplement on Cytokine Production from Allergic Rhinitis Patients

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ABSTRACT  Spirulina represents a blue-green alga that is widely produced and commercialized as a dietary supplement for modulating immune functions, as well as ameliorating a variety of diseases. We have previously shown that the in vitro culture of Spirulina with human peripheral blood mononuclear cells (PBMCs) modulated the production of cytokines. In the present study, we evaluated the impact of a Spirulina-based dietary supplement (Earthrise Nutritional, Inc., Irvine, CA) on patients with allergic rhinitis by assessing the production of cytokines (interleukin-1β, interferon (IFN)-γ, and IL-2) critical in regulating immunoglobulin E-mediated allergy. In a randomized double-blinded crossover study versus placebo, allergic individuals were fed daily with either placebo or Spirulina, at 1,000 mg or 2,000 mg, for 12 weeks. PBMCs isolated before and after the Spirulina feeding were stimulated with phytohemagglutinin (PHA) prior to determining the levels of cytokine from cell culture supernatants. Although Spirulina seemed to be ineffective at modulating the secretion of Th1 cytokines (IFN-γ and IL-2), we discovered that Spirulina, administered at 2,000 mg/day, significantly reduced IL-4 levels by 32% from PHA-stimulated cells. These results indicate that Spirulina can modulate the Th profile in patients with allergic rhinitis by suppressing the differentiation of Th2 cells mediated, in part, by inhibiting the production of IL-4. To our knowledge, this is the first human feeding study that demonstrates the protective effects of Spirulina towards allergic rhinitis.

KEY WORDS: • antioxidants  • blue-green algae  • interleukin-4  • Spirulina

INTRODUCTION

Spirulina is a type of cyanobacteria with a history of human consumption dating as far back as the 16th Century, where it has been reported that Aztecs harvested algal biomass resembling Spirulina to be consumed as part of their diet.1 Spirulina is a filamentous blue-green alga that apparently flourishes in alkaline lakes with an extremely high pH.2 This cyanobacterium can potentially represent an important staple in the human diet due to its overall nutritional qualities, which includes a high content of protein (60–71% according to strain), phenolic acids, tocopherols, -carotenes, and -linolenic acid.1 In addition to its nutritional values, Spirulina lacks cellulose cell walls and therefore does not require chemical or physical processing in order to become digestible.1 Moreover, Spirulina is relatively easy to cultivate, thereby sparking the early interest in it as a commercial food supplement with potential therapeutic health benefits.

Not only can the consumption of Spirulina be healthy by supplying proteins and other nutrients, but a number of reports have suggested Spirulina to be an effective immune modulator. It has been well documented that Spirulina exhibited anti-inflammatory properties, in particular, by inhibiting the release of histamine from mast cell-mediated allergic reactions.3-5 The active ingredient found in Spirulina responsible for its anti-inflammatory activities is C-phycocyanin, a pigment commonly found in blue-green algae.6-9 It is known that C-phycocyanin can selectively inhibit the activity of cyclooxygenase-2, a critical enzyme in the biosynthesis of prostaglandins.9 Because of its chemical structure, C-phycocyanin also displays antioxidative and free radical scavenging properties, which may contribute, at least in part, to its anti-inflammatory activities.6,10-13 Additional studies have also indicated that Spirulina exposure enhances the phagocytic functions of macrophages in cats,14 chickens,15 and channel catfish.16 Furthermore, dietary Spirulina was shown to have protective effects toward orally induced food allergy in mice by preventing the increase of immunoglobulin (Ig) E, while increasing the IgA antibody level to protect against allergic reactions.17 More recently, we have shown that in vitro culture of Spirulina extract was effective in modulating the secretion of cytokines [interleukin (IL)-1β, IL-4, and interferon (IFN)-γ] from human peripheral blood mononuclear cells (PBMCs).18 In continuation of such promising studies, with particular emphasis on inflammatory-mediated diseases such as al-
lergy, the present study evaluates the impact of dietary *Spirulina* on cytokine production from patients with allergic rhinitis. In a randomized double-blinded crossover study versus placebo, subjects diagnosed with clinical allergic rhinitis were fed daily with either *Spirulina* (at 1,000 or 2,000 mg) or placebo for 12 weeks. PBMCs, collected from the subjects before and after *Spirulina* feeding, were cultured in the presence and absence of phytohemagglutinin (PHA) for 48 hours, after which cytokine (IL-4, IFN-γ, and IL-2) levels were determined from cell culture supernatants.

**MATERIALS AND METHODS**

**Randomized double-blinded crossover versus placebo study**

A total of 36 patients ranging between 18 to 55 years of age with a clinical history of allergic rhinitis, but no other health problems, were enrolled in this study. The patients were equally divided into three groups, each consuming either placebo (Group D) or a proprietary, *Spirulina*-based dietary supplement of Earthrise Nutritional, Inc. (Irvine, CA) at doses of 1,000 mg/day (Group C) or 2,000 mg/day (Group U) for 12 weeks. All subjects were instructed to ingest four capsules per day. Therefore, the group who received the *Spirulina* supplement at 1,000 mg/day (Group C) was also taking inert placebo as part of their dosage. The placebo was a color-matched capsule containing inert material. It should be noted that two subjects from group U dropped out of the study, leaving a group size of 10. This study was approved by the Institutional Review Board at the University of California at Davis (Protocol number 200210882-1).

**PBMC isolation**

Peripheral blood was collected at week 0 pre-feeding and week 12 post-feeding into sodium citrate-containing tubes. The blood was diluted 1:1 with Hank's Balanced Salt Solution (HBSS; Invitrogen, Carlsbad, CA) without calcium chloride, magnesium chloride, or magnesium sulfate. The diluted blood was then layered over a Histopaque®-1077 gradient (Sigma, St. Louis, MO) and centrifuged at 500 g for 30 minutes at room temperature. PBMCs were harvested from the interface layer, washed twice with HBSS, and then counted. The cells were resuspended in RPMI-1640 (Invitrogen) containing 10% fetal bovine serum and supplemented with 0.1% of a 50 mg/mL gentamicin solution (Invitrogen). PBMC concentration was adjusted to 2 × 10^6 viable cells/mL after estimation of viability by trypan blue exclusion assay. Viability was consistently greater than 96%.

**Culture of PBMCs**

In 48-well plates, a cell suspension containing 1.0 × 10^6 PBMCs was cultured in the presence or absence of PHA (25 g/mL) at 37°C with 5% CO2. Following a 48-hour incubation, the supernatant fractions were harvested for enzyme-linked immunosorbent assay (ELISA) analysis. All treatments were performed in duplicate.

**ELISA analysis of cytokines**

Levels of IL-4, IFN-γ, and IL-2 were measured in supernatants from PHA-stimulated and unstimulated PBMCs. For quantitation of secretory IL-4, a High Sensitivity Quantikine Human IL-4 ELISA kit (R&D Systems, Minneapolis, MN) was used with a detection limit of 0.25 pg/mL. Standard ELISA kits used to measure levels of IFN-γ and IL-2 (R&D Systems, Minneapolis, MN) had a detection limit of 15.6 pg/mL.

**Statistics**

Results obtained following the *Spirulina* feeding (week 12) were compared with baseline (week 0) by Student’s paired *t* test with a two-tailed *P* value. Significance was taken as *P* < 0.05.

**RESULTS**

**Cytokine production from unstimulated PBMCs**

PBMCs cultured for 48 hours under non-stimulatory conditions (i.e., media only) did not produce any detectable levels of IL-4, IFN-γ, and IL-2 in any groups.

**Cytokine production from PHA-stimulated PBMCs**

When cells were cultured in the presence of a T-cell mitogen, PHA was able to induce measurable levels of cyto-

![IFN-γ concentration](image-url)
only been within the last decade that the scientific community has corroborated some of the alleged health benefits of Spirulina. In particular, it is well documented that this cyanobacterium exhibits anti-inflammatory properties, and therefore can potentially alleviate certain diseases mediated by inflammation, such as colitis and allergy. In the present study, we examined the therapeutic application of dietary Spirulina towards allergic rhinitis by measuring production of cytokines important in regulating IgE-mediated allergy.

Allergic rhinitis is thought to be initiated by an allergen breaching the mucosal epithelium, where antigen-presenting cells, particularly dendritic cells, can take up, process, and present allergen peptides to type-2 T helper (Th2) cells. This cellular interaction stimulates Th2 cells to release cytokines that promote the IgE production, as well as the growth and activation of eosinophils, mast cells, and basophils. In particular, IL-4 is a critical cytokine involved in all aspects of inducing type I hypersensitivity reactions. It is well known that IL-4 can effectively promote IgE production from B cells, which can sensitize an individual to a particular allergen by binding to high-affinity FcRI present on mast cells. Therefore, subsequent allergen binding to IgE on the surface of mast cells can culminate in the release of inflammatory mediators, initiating symptoms characteristic of allergic rhinitis. Hence, aberrant production of IL-4 has been implicated in allergy for it favors Th2 differentiation in correlation with humoral-mediated immunity. In con-

**DISCUSSION**

For thousands of years, humans have been incorporating Spirulina as part of their diet on the assumption that this prehistoric plant can combat various ailments. Yet, it has...
The Th2 cells can secrete IL-2 and IFN-α to antagonize the cytokine milieu that promotes allergic inflammation. IFN-α is a macrophage-activating cytokine that promotes Th1-biased responses associated with cell-mediated immunity. Consequently, the balance of Th1/Th2 type cytokines is critical in determining whether an immune response is to be dominated by macrophage activation or by antibody production, particularly IgE, which mediates symptoms of allergic rhinitis.

In the present double-blinded crossover study, we have shown that allergic patients consuming 2,000 mg of Spirulina daily can reduce the production of IL-4 from PHA-stimulated PBMCs by 32%. According to several reports claiming that Spirulina-fed animals can inhibit mast cell function, it is now conceivable such activity may, in part, have resulted from reduced levels of IL-4. Although we cannot confirm that such inhibition observed is dose-dependent, it may be possible that increasing the daily regimen above 2,000 mg may also enhance the suppressing capability of Spirulina.

In our previous investigation, we reported that in vitro culture of resting and PHA-stimulated PBMCs with Spirulina extract significantly increased the levels of IL-4. However, it is important to consider that the subjects participating in our prior study were healthy individuals that did not suffer from allergic rhinitis. Furthermore, the baseline value for IL-4 from PHA-stimulated PBMC cultures of all allergic patients (22.1 ± 2.3 pg/mL, n = 34) in our current study (prior to the Spirulina feed) is 69.5% higher than similar cultures observed with healthy individuals (13.1 ± 6.9 pg/mL, n = 12; values taken from Mao et al.). Hence, it is conceivable that the Spirulina feed modulated the Th profile in allergic individuals either by preventing the differentiation of Th2 cells or by enhancing the population of Th1 cells to overcome the effects of Th2 cells. Since we did not observe any significant difference in IFN-α and IL-2 (both Th1-type cytokines) production following 12 weeks, it is most likely that Spirulina exerted its effects by suppressing the pathway leading to Th2-committed cells mediated, in part, by inhibiting the secretion of IL-4.

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REFERENCES