

**REPLICATION IN IMAGING GENETICS:
THE CASE OF THREAT-RELATED AMYGDALA REACTIVITY**

Supplementary Information

Supplementary Methods

Inclusion/Exclusion Criteria of Previous Studies. Because our genetic data is restricted to SNPs, we excluded 37 studies, which tested other types of polymorphisms (e.g., variable number tandem repeats). We also had to exclude 6 studies, which used tasks or contrasts that were not related to threat (e.g., gambling task or sad facial expressions) and were thus incomparable to our fMRI data. Two additional studies were excluded because they were based on clinical samples not well represented in our dataset (i.e., schizophrenia and posttraumatic stress disorder). One study did not find a significant association between 23 tagging-SNPs covering the oxytocin receptor gene and amygdala reactivity (1) and was therefore not included (2 of the 23 SNPs were represented by included studies). As several studies reported on the same SNPs, there were 37 unique associations we attempted to replicate (Table S1).

Inclusion/Exclusion Criteria of Participants. All participants were free of the following study exclusions: 1) medical diagnoses of cancer, stroke, diabetes requiring insulin treatment, chronic kidney or liver disease, or lifetime history of psychotic symptoms; 2) use of psychotropic, glucocorticoid, or hypolipidemic medication; and 3) conditions affecting cerebral blood flow and metabolism (e.g., hypertension). Diagnosis of any current DSM-IV Axis I disorder or select Axis II disorders (antisocial personality disorder and borderline personality disorder), assessed with the electronic Mini International Neuropsychiatric Interview (2) and Structured Clinical Interview for the DSM-IV subtests (3), respectively, were not an exclusion criterion, as the Duke Neurogenetics Study seeks to establish broad variability in multiple behavioral phenotypes related to psychopathology. However, no

individuals, regardless of diagnosis, were taking any psychoactive medication during or at least 14 days prior to their participation. Of the 1117 participants used in our analyses, 236 individuals had at least one DSM-IV diagnosis, including 121 with alcohol use disorders, 43 with non-alcohol substance use disorders, 64 with major depressive disorders, 32 with bipolar disorders, 24 with panic disorder (no agoraphobia), 21 with panic disorder including agoraphobia, 11 with social anxiety disorder, 20 with generalized anxiety disorder, 13 with obsessive compulsive disorder, 10 with eating disorders, and 2 with posttraumatic stress disorder.

Genotype Imputation. Genotype imputation was performed on all DNS participants with genome-wide chip data using the prephasing/imputation stepwise approach implemented in SHAPEIT/IMPUTE2 (4, 5). Imputation was run separately for participants genotyped on the Illumina HumanOmniExpress (n = 685) and the Illumina HumanOmniExpress-24 (n = 432) arrays using biallelic SNPs only, the default value for effective size of the population (20,000), and chunk sizes of 3 Mb and 5 Mb for the respective arrays. Within each array batch, genotyped SNPs used for imputation were required to have missingness $<.02$, Hardy-Weinberg equilibrium $p > 10^{-6}$, and minor allele frequency $>.01$. The imputation reference set consisted of 2504 phased haplotypes from the full 1000 Genomes Project Phase 3 data set (May 2013, >70 million variants, release "v5a"). Imputed SNPs were retained if they had high imputation quality (Info $>.9$), low missingness ($<5\%$), and minor allele frequency $>.01$. A proxy was used if the target SNP was neither directly genotyped nor imputable. Proxy SNPs were found with the "LDproxy" tool in "<https://analysistools.nci.nih.gov/LDlink/>" based on the "Utah residents from north and west Europe" population. All proxies were in perfect linkage

disequilibrium with the target SNP ($D'=1$, $R^2=1$), except for rs853180 ($D'=.95$, $R^2=.89$). Details for the derivation of each target SNP are provided in Table S2. The alleles of six of the original SNPs were C/G or A/T, and therefore the target allele of the proxy was also matched based on frequency.

fMRI Task. The task version used in the DNS consisted of four blocks of a face-processing task interleaved with five blocks of a sensorimotor control task. During task blocks, participants viewed six trios of either fearful, angry, surprised, or neutral faces and matched one of two faces (bottom) identical to a target face (top). Each trial in the face-matching blocks lasted for 4 seconds with a variable interstimulus interval of 2–6 seconds (mean = 4 seconds). In the control blocks, each of six shape trios was presented for 4 seconds with a fixed interstimulus interval of 2 seconds. Each block began with a 2 second instruction screen (i.e., “Match Faces” or “Match Shapes”) resulting in a total task time of 390 seconds.

BOLD fMRI Data Acquisition. Each participant was scanned using one of two identical research-dedicated GE MR750 3T scanner equipped with high-power high-duty-cycle 50-mT/m gradients at 200 T/m/s slew rate, and an eight-channel head coil for parallel imaging at high bandwidth up to 1MHz at the Duke-UNC Brain Imaging and Analysis Center.. A semi-automated high-order shimming program was used to ensure global field homogeneity. A series of 34 interleaved axial functional slices aligned with the anterior commissure-posterior commissure plane were acquired for full-brain coverage using an inverse-spiral pulse sequence to reduce susceptibility artifacts (TR/TE/flip angle=2000 ms/30 ms/60; FOV=240mm; 3.75×3.75×4mm voxels; interslice skip=0). Four initial radiofrequency excitations were performed (and discarded) to achieve steady-state equilibrium. To allow for

spatial registration of each participant's data to a standard coordinate system, high-resolution three-dimensional T1-weighted structural images were obtained in 162 axial slices using a 3D Ax FSPGR BRAVO sequence (TR/TE/flip angle = 8.148 ms / 3.22 ms / 12°; voxel size=0.9375x0.9375x1mm; FOV=240mm; interslice skip=0; total scan time = 4 min and 13 s). In addition, high-resolution structural images were acquired in 34 axial slices coplanar with the functional scans and used for spatial registration for participants without Ax FSPGR BRAVO images (TR/TE/flip angle=7.7 s/3.0 ms/12; voxel size=0.9x0.9x4mm; FOV=240mm, interslice skip=0).

BOLD fMRI Data Pre-Processing. Anatomical images for each subject were skull-stripped, intensity-normalized, and nonlinearly warped to a study-specific average template in a standard stereotactic space (Montreal Neurological Institute template) using ANTs (6). BOLD time series for each subject were processed in AFNI (7). Images for each subject were despiked, slice-time-corrected, realigned to the first volume in the time series to correct for head motion, coregistered to the anatomical image using FSL's Boundary Based Registration (8), spatially normalized into MNI space using the non-linear warp from the anatomical image, resampled to 2mm isotropic voxels, and smoothed to minimize noise and residual difference in gyral anatomy with a Gaussian filter, set at 6-mm full-width at half-maximum. All transformations were concatenated so that a single interpolation was performed. Voxel-wise signal intensities were scaled to yield a time series mean of 100 for each voxel. Volumes exceeding 0.5mm frame-wise displacement or 2.5 standardized DVARS (9, 10) were censored from the GLM.

The AFNI program 3dREMLfit (7) was used to fit a general linear model for first-level fMRI data analyses. Following preprocessing, linear contrasts employing canonical

hemodynamic response functions were used to estimate expression specific effects (e.g., angry expression blocks > control blocks) as well as a general threat effect (i.e., anger and fear expression blocks > control blocks) for each individual. These Individual contrast images were then used in second-level random effects models in SPM12 (<http://www.fil.ion.ucl.ac.uk/spm>) accounting for scan-to-scan and participant-to-participant variability to determine mean condition-specific regional responses using one-sample t-tests. A statistical threshold of $p < 0.05$, family-wise error corrected across our anatomical amygdala regions of interest (defined using a high-resolution templated generated from 168 Human Connectome Project datasets 11) was applied to the contrasts. Specific region of interest (ROI) analyses for the combined Central and Medial Nuclei of the amygdala were also conducted as two of the studies (12, 13) reported associations for this amygdala subregion only.

fMRI Quality Assurance Criteria. Quality control criteria for inclusion of a participant's imaging data were: >5 volumes for each condition of interest retained after censoring for FD and DVARS and sufficient temporal SNR within the bilateral amygdala, defined as greater than 3 standard deviations below the mean of this value across subjects. The amygdala was defined using a high-resolution templated generated from 168 Human Connectome Project datasets (11). Additionally, data were only included in further analyses if the participant demonstrated sufficient engagement with the task, defined as achieving at least 75% accuracy during the face matching condition.

Table S1. Studies reporting on an association between a genetic variation and threat-related amygdala reactivity.

Included/Excluded	Reason for exclusion	Gene	Study
Excluded	Not a SNP	5HTTLPR	Oathes et al., 2015 (14)
Excluded	Not a SNP	5HTTLPR	Shneck et al., 2016 (15)
Excluded	Not a SNP	5HTTLPR	Stoeckel et al., 2015 (16)
Excluded	Not a SNP	5HTTLPR	Bastiaansen et al., 2014 (17)
Excluded	Not a SNP	5HTTLPR	Klumpp et al., 2014 (18)
Excluded	Not a SNP	5HTTLPR	Costafreda et al., 2013 (19)
Excluded	Not a SNP	5HTTLPR	Klucken et al., 2013 (20)
Excluded	Not a SNP	5HTTLPR	O'Nions et al., 2011 (21)
Excluded	Not a SNP	5HTTLPR	Morey et al., 2011 (22)
Excluded	Not a SNP	5HTTLPR	Lonsdorf et al., 2011 (23)
Excluded	Not a SNP	5HTTLPR	Lemogne et al., 2011 (24)
Excluded	Not a SNP	5HTTLPR	Kobiella et al., 2011 (25)
Excluded	Not a SNP	5HTTLPR	Furman et al., 2010 (26)
Excluded	Not a SNP	5HTTLPR	Dannlowski et al., 2010 (27)
Excluded	Not a SNP	5HTTLPR	Gillihan et al., 2011 (28)
Excluded	Not a SNP	5HTTLPR	Lau et al., 2009 (29)
Excluded	Not a SNP	5HTTLPR	Rao et al., 2007 (30)
Excluded	Not a SNP	5HTTLPR	Dannlowski et al., 2008 (31)
Excluded	Not a SNP	5HTTLPR	Heinz et al., 2007 (32)
Excluded	Not a SNP	5HTTLPR	Bertolino et al., 2005 (33)
Excluded	Not a SNP	5HTTLPR	Hariri et al., 2005 (34)
Excluded	Not a SNP	5HTTLPR	Hariri et al., 2002 (35)
Excluded	Not a SNP	5HTTLPR	Lee and Ham 2008 (36)
Excluded	Not a SNP	5HTTLPR	Smolka et al., 2007 (37)
Excluded	Not a SNP	MAOA	Lee & Ham 2008 (38)
Excluded	Not a SNP	NOS1	Kuhn et al., 2016 (39)
Excluded	Not a SNP	MAOA	Meyer-Lindenberg et al., 2006 (40)
Excluded	Not a SNP	MAOA	Denson et al., 2014 (41)
Excluded	Not a SNP	DAT1/SLC6A3	Wetherill et al., 2014 (42)
Excluded	Not a SNP	DAT1/SLC6A3	Bergman et al., 2014 (43)
Excluded	Not a SNP	AR	Manuck et al., 2010 (44)
Excluded	Not a SNP	ADRA2B	Rasch et al. 2009 (45)

Included/Excluded	Reason for exclusion	Gene	Study
Excluded	Not a SNP	ADRA2B	Cousijn <i>et al.</i> , 2010 (46)
Excluded	Not a SNP	ADRA2B	Urner <i>et al.</i> , 2011 (47)
Excluded	Not a SNP	ADRA2B	Li <i>et al.</i> , 2015 (48)
Excluded	Not a SNP	AVPR1A	Meyer-Lindenberg <i>et al.</i> , 2009 (49)
Excluded	Not a SNP	FMR1	Hessl <i>et al.</i> , 2007 (50)
Excluded	Language task	GWAS bipolar disorder score	Whalley <i>et al.</i> , 2012 (51)
Excluded	Monetary task	ODZ4	Heinrich <i>et al.</i> , 2013 (52)
Excluded	Alcohol related task	GATA4	Jorde <i>et al.</i> , 2014 (53)
Excluded	Sad > Neutral faces	HTR2A	Lee & Ham., 2008 (36)
Excluded	Sad > Neutral faces	TPH1	Lee <i>et al.</i> , 2009 (54)
Excluded	Gambling videos	DBH	Yang <i>et al.</i> 2016 (55)
Excluded	Only significant in SCZ	OXTR	Haram <i>et al.</i> , 2016 (56)
Excluded	Null finding	OXTR	Loth <i>et al.</i> , 2014 (1)
Excluded	PTSD risk sample	ADCYAP1R1	Stevens <i>et al.</i> , 2014 (57)
Included		PCDH17	Chang <i>et al.</i> , 2017 (58)
Excluded	Fakra <i>et al.</i> , 2009 (59) used as target study	HTR1A	Domschke <i>et al.</i> , 2006 (60) Panic disorder patients.
Excluded	Fakra <i>et al.</i> , 2009 (59) used as target study	HTR1A	Straube <i>et al.</i> , 2014 (61) A clinical sample of panic disorder patients with agoraphobia.
Included		HTR1A	Fakra <i>et al.</i> , 2009 (59)
Included		NPSR1	Dannlowski <i>et al.</i> , 2011 (27)
Included		RGS2	Smoller <i>et al.</i> , 2008 (62)
Included		ASIC1/ACCN2	Smoller <i>et al.</i> , 2014 (63)
Excluded	Hariri <i>et al.</i> , 2009 (64) used as target study	FAAH	Gunduz-Cinar <i>et al.</i> , 2013 (65)
Included		FAAH	Hariri <i>et al.</i> , 2009 (64)
Included		NR3C2	Bogdan <i>et al.</i> , 2012 (66)
Included		PET-1	Wellman <i>et al.</i> , 2013 (67)
Included		CHT1	Neumann <i>et al.</i> , 2006 (68)
Included		FKBP5	Holz <i>et al.</i> , 2015 (69)
Included		OXTR	Tost <i>et al.</i> , 2010 (70)
Included		IFN- γ	Redlich <i>et al.</i> , 2015 (71)
Included		ADCY7	Joeyen-Waldorf <i>et al.</i> , 2012 (72)

Included/Excluded	Reason for exclusion	Gene	Study
Included		GTF2I	Swartz et al., 2015 (73)
Included		VMAT1	Lohoff et al., 2014 (74)
Included		FREM3	Nikolova et al., 2015 (75)
Included		FREM3	Nikolova et al., 2015 (75)
Included		CACNA1C	Tesli et al., 2013 (76)
Excluded	Tesli et al., 2013 (76) used as target study	CACNA1C	Sumner et al., 2015 (77) 58 adolescents and young adults viewing negative > neutral images, AA higher activation.
Excluded	Tesli et al., 2013 (76) used as target study	CACNA1C	Jogia et al., 2011 (78) 116 healthy and BD patients; A carriers higher activation; fearful > neutral.
Excluded	Tesli et al., 2013 (76) used as target study	CACNA1C	Wessa et al., 2010 (79) Reward task, 64 participants, A allele associated with higher activation in right amygdala.
Included		19 SNPs related to GR	Arloth et al., 2015 (12)
Included		PHOX2B	Ousdal et al., 2012 (80)
Included		OXTR	Waller et al., 2016 (81)
Included		CNIH3	Nelson et al., 2016 (82)
Included		IL-18	Swartz et al., 2016 (13)
Included		COMT	Domschke et al., 2012 (83)
Excluded	Domschke et al., 2012 used as target study	COMT	Lonsdorf et al., 2011 (23) 54 participants, 29 females, age 24, passively viewing two white male angry faces, white fixation cross, angry > control, left amygdala, Met/Met higher.
Excluded	Domschke et al., 2012 used as target study	COMT	Kempton et al., 2009 (84) 34 females, sample of 74, 1.5T, fear facial expression changing intensity, extracted values, higher in left amygdala with Val/Val.
Excluded	Domschke et al., 2012 used as target study	COMT	Smolka et al., 2005 (85) 35 subjects, 26 males, age 41, 1.5T, Caucasians, affectively unpleasant, pleasant, and neutral pictures, unpleasant > neutral, right amygdala, Met carriers.
Excluded	Domschke et al., 2012 used as target study	COMT	Drabant et al., 2006 (86) 101 subjects, 50 females, age 30, 3T, face matching task, fearful and angry, did not find link.
Excluded	Domschke et al., 2012 used as target study	COMT	Klucken et al., 2015 (87) 80 subjects, appetitive conditioning, erotic pictures as UCS, 1.5T, CS+ > CS-, left amygdala, Val/Val.
Excluded	Domschke et al., 2012 used as target study	COMT	Lelli-Chiesa et al., 2011 (88) sample of 40 BD, 25 healthy relatives, 22 relatives with MDD or anxiety, and 50 healthy controls, 1.5T, extracted values, sad face intensity and neutral, sad > neutral, Val carriers, no effect of group.
Excluded	Domschke et al., 2012 used as target study	COMT	Rasch et al., 2010 (89) 56 subjects, 15 males, age 24, positive, negative, and neutral pictures, 3T, unpleasant > neutral, Met higher in right amygdala.

Included/Excluded	Reason for exclusion	Gene	Study
Excluded	Domschke et al., 2012 used as target study	COMT	Domschke et al., 2008 (90) 20 patients with panic disorder, 3T, tested with various facial expression, found with fear > grey rectangle - the control stimulus, right amygdala Val carriers.
Excluded	Domschke et al., 2012 used as target study	COMT	Opmeer et al., 2014 (91) 125 participants, 28 healthy and rest MDD, mostly females, age 38, gender judgment task on various emotional faces, scrambled face and button press as control, 3T, positive expressions > control, left amygdala, Met carriers, but most of it was the hippocampus.
Excluded	Domschke et al., 2012 used as target study	COMT	Williams et al., 2010 (92) 46 participants, 21 females. Masked and unmasked fear, happy, and neutral, 1.5T, fear > neutral unmasked, met carriers, bilateral.
Excluded	Domschke et al., 2012 used as target study	COMT	Smolka et al., 2007 (37) 48 subjects all males, age 41, Caucasians, 1.5T, unpleasant, pleasant, and neutral pictures, unpleasant > neutral, Met carriers.
Excluded	Kilpatrick et al., 2011 (93) used as target study	HTR3A	Iidaka et al., 2005 (94) Japanese sample; 26 people.
Included		HTR3A	Kilpatrick et al., 2011 (93)
Included		DRD2/ANKK1	Lee et al., 2011 (95)
Excluded	Canli et al., 2005(96)used as target study	TPH2	Brown et al., 2005 (97)
Included		TPH2	Canli et al., 2005 (96)
Included		DRD2	Blasi et al., 2009 (98)
Included		PCLO	Woudstra et al., 2012 (99)
Included		IL1B	Baune et al.,2010 (100)
Included		DOK5	Liu et al., 2010 (101)
Included		NPY	Zhou et al., 2008 (102)
Included		NPY	Domschke et al., 2010 (103)
Included		BDNF	Soliman et al., 2010 (104)
Excluded	Soliman et al., 2010 (104) used as target study	BDNF	Montag et al., 2008 (105) Sample of 37 women; mean age 23; viewing emotional pictures.
Excluded	Soliman et al., 2010 (104) used as target study	BDNF	Lau et al., 2010 (106) Greater response only in MDD and anxiety adolescent patients.
Excluded	Soliman et al., 2010 (104) used as target study	BDNF	Gasic et al., 2009 (107) Val/Val 29 Caucasians.
Included		NR3C1	Ridder et al., 2012 (108)
Included		NR3C1	Ridder et al., 2012 (108)
Included		NR3C1	Ridder et al., 2012 (108)

Included/Excluded	Reason for exclusion	Gene	Study
Included		CRHR1	Weber et al., 2016 (109)
Included		OXTR	Marusak et al., 2015 (110)

Table S2. Summary of 37 candidate gene association studies of threat-related amygdala reactivity

Target Study	Demographics	Ethnicity	Gene	Gene product	Polymorphism	Functionality
Chang et al., 2017 (58)	N=297 (163 females), age: 33.5	European descent	PCDH17	Cadherin protein, associated with cell to cell interactions in the brain.	rs9537793	G allele associated with higher mRNA levels (58)
Fakra et al., 2009 (111)	N=89 (44 females), age: 45.74	Caucasian	HTR1A	Serotonin receptor 1A	rs6295 (used proxy rs358527)	G allele leads to higher expression levels of the 5-HT1A autoreceptor (112)
Dannowski et al., 2011 (27)	N=72 (41 females), age: 36.77	European	NPSR1	Neuropeptide S receptor 1. Has an anxiolytic effect. Possibly affects the neurotransmission of serotonin, dopamine, and noradrenaline	rs324981 (used proxy rs324987)	the T-allele (Ile) leads to increased NPSR expression (113, 114)
Smoller et al., 2008 (62)	N=55 (41 females), age: 19.52	Missing (but controlled for ancestry based on informative marker clusters)	RGS2	Regulator of G protein signaling 2. Accelerates deactivation of G proteins to reduce G protein-coupled receptor signaling. Related to anxiety.	rs4606 (used proxy rs3767488)	G allele is associated with low RGS2 expression and increased G-protein-coupled signaling (115)
Smoller et al., 2014 (63)	N=103 (54 females), age: 20.37	Caucasians	ACCN2	Amiloride-sensitive cation channel 2. A sodium/calcium exchanger, expressed in the brain and kidney, antihypertensive drug Amiloride binds to it in the kidney.	rs10875995	Unknown
Hariri et al., 2009 (64)	N=82 (39 females), age: 44.85	Caucasian	FAAH	Fatty acid amide hydrolase. A key enzyme in regulating endocannabinoid (eCB) signaling.	rs324420	A allele associated with reduced expression of FAAH and elevations in circulating levels of the eCB anandamide (65)
Bogdan et al., 2012 (66)	N=279 (140 females), age: 13.59	Caucasian, Hispanic, and other	NR3C2	Nuclear Receptor Subfamily 3 Group C Member 2. A receptor with equal affinity to mineralocorticoids and glucocorticoids. Involved in the negative feedback loop of the HPA axis.	rs5522	The G allele has been associated with lower efficiency in using cortisol as ligand (116, 117)
Neumann et al., 2006 (68)	N=32 (14 females), age: 41.13	European	CHT1	Human choline transporter gene. Carries choline into acetylcholine-synthesizing neurons.	rs333229 (imputed)	Unknown
Holz et al., 2015 (69)	N=153 (87 females), age: 25	European	FKBP5	FK506 binding protein 5. Higher levels of the protein coded by this	rs1360780	T allele associated with higher FKBP5 expression and impaired

Target Study	Demographics	Ethnicity	Gene	Gene product	Polymorphism	Functionality
				gene lead to impaired HPA axis negative feedback.		recovery of cortisol levels in response to stress (118, 119)
Tost et al., 2010 (70)	N=228 (126 females), age=31.9	Caucasian	OXR	Oxytocin receptor	rs53576	Unknown
Redlich et al., 2015 (71)	N=337 (218 females), age:36.46	Caucasian	IFN- γ	Interferon γ . A cytokine that is critical for innate and adaptive immune response	rs1861494	C allele introduces a new CpG dinucleotide sequence that serves as an epigenetic target for DNA methylation, leading to reduced gene expression (120)
Joeyen-Waldorf et al., 2012 (72)	Sample 1: N=82 (46 females), age: 44.76; Sample 2: N=98 (40 females), age: 40.53	Caucasian	ADCY7	Adenylate cyclase 7. A membrane-bound adenylyl cyclase that catalyses the formation of cyclic AMP from ATP.	rs1064448	Unknown
Tesli et al., 2013 (76)	N=250 (66 BD, 61 SZ, 114 females), age:34.4	Northern European	CACNA1C	a1c subunit of the L-type voltage-gated calcium channel. Implicated in synaptic plasticity and neuronal survival.	rs1006737	Unknown
Ousdal et al., 2012 (80) ^{^^}	N=224 (51 SZ, 64 BD, and 12 with 'other psychosis', 109 females), age: 32.1	Missing (but mention that individuals with a calculated ancestry different from the majority of the TOP sample were removed)	PHOX2B	Paired-like homeobox 2b. A homeodomain transcription factor. Regulates the expression of enzymes necessary for the synthesis of several monoamines and is essential for the development of the autonomic nervous system	rs10014254 (imputed)	Unknown
Blasi et al., 2009 (98)	N=134 (84 females), age: 26.4	Caucasians	DRD2	Dopamine receptor D2	rs1076560	GG genotype associated with relatively greater expression of DRD2 short (autoreceptor) mRNA in prefrontal cortex and in striatum (121)
Kilpatrick et al., 2011 (93)	N=55 (26 IBS, all females), age: 32.54	Missing	HTR3A	Serotonin receptor 3A	rs1062613	T allele leads to increased HTR3A expression (122)
Lee et al., 2011 (95)	N=45 (all females), age: 23.15	Asian	DRD2/ANKK1	Ankyrin repeat and kinase domain containing 1. While the SNP is located in the ANKK1 gene it is thought to affect DRD2 expression	rs1800497	A1 (T) allele predicts lower availability of DRD2 in the striatum (123), but in Zhang et al. (121) a direct effect of this variant was not found and it was postulated that previous found

Target Study	Demographics	Ethnicity	Gene	Gene product	Polymorphism	Functionality
Canli et al., 2005 (96)	N=29 (18 females), age: 22.8	Missing (but mention that ethnicity did not differ according to genotype)	TPH2	lisoenzyme of tryptophan hydroxylase, the rate-limiting enzyme in the biosynthesis of serotonin	rs4570625	Unknown effects were due to an LD with rs1076560
Woudstra et al., 2012 (99)	N=126 (96 MDD, 78 females), age:38.1	Western European	PCLO	Piccolo Presynaptic Cytomatrix Protein. Involved in monoamine neurotransmission. Part of the presynaptic cytoskeletal matrix, which is involved in establishing active synaptic zones and in synaptic vesicle trafficking.	rs2522833	Unknown
Zhou et al., 2008 (102)	N=71 (40 females), age: 44.4	91% Caucasians	NPY	neuropeptide Y. Involved in anxiety and stress. Has anxiolytic effects.	Haplotypes of 6 SNPs (rs3037354, rs17149106, rs16147, rs16139, rs5573, rs5574)	The various haplotypes are associated with mRNA expression and plasma levels(102)
Domschke et al., 2010 (103)	N=35 (all MDDs, 24 females), age: 37.3	Northern European	NPY		rs16147	C allele associated with decreased expression (102)
Baune et al.,2010 (100)	N=32 (all MDDs, 22 females), age:37.8	Caucasian	IL-1 β	Interleukin 1 β . Proinflammatory cytokine.	rs16944	Mixed findings regarding which allele is associated with higher expression. May be functional in a haplotype (124)
^Wellman et al., 2013(67)	N=89 (50 females), age: 19.55	Asians	PET-1	Pheochromocytoma 12 ETS factor-1; FEV-1. A transcription factor essential for differentiation and forebrain targeting of serotonin neurons	rs860573	Unknown
^Nelson et al., 2016 (82)	N=312 (161 females), age: 19.71	European American	CNIH3	Cornichon family AMPA receptor auxiliary protein 3. Regulates the trafficking and gating properties of AMPA-selective glutamate receptors	rs10799590	Unknown
^Waller et al., 2016 (81)	N=312 (213 females), age: 19.71	Non Hispanic Caucasians	OXTR	Oxytocin receptor	rs1042778	Unknown
^Swartz et al., 2015 (73)	N=808 (454 females), age: 19.71	Caucasian, African	GTF2I	General transcription factor Iii. This locus, along with several other neighboring genes, is deleted in	rs13227433 (imputed)	Unknown

Target Study	Demographics	Ethnicity	Gene	Gene product	Polymorphism	Functionality
		American, and Asian		Williams-Beuren syndrome, which is characterized by hypersociability and increased approach to strangers.		
[^] Swartz et al., 2016 (13)	N=448 (261 females), age: 19.71	Non Hispanic Caucasians	IL-18	Interleukin 18. Proinflammatory cytokine	rs187238 (used proxy rs795467) and rs1946518	Haplotype comprising the major C risk allele of rs187238 and the major G risk allele of rs1946518 associated with increased expression of IL-18 mRNA (125) and plasma IL-18 (126).
[^] Lohoff et al., 2014 (74)	N=298 (173 females), age: 19.63	Caucasian, Asian, African American, Latino, other	VMAT1	Vesicular monoamine transporter 1. Embedded in synaptic vesicles and serves to transfer monoamines, such as norepinephrine, epinephrine, dopamine, and serotonin, between the cytosol and synaptic vesicles.	rs1390938 (imputed)	A allele leads to increased monoamine transport into presynaptic vesicles (74)
[^] Nikolova et al., 2015 (75)	N=365 (192 females), age: 19.71	Non Hispanic Caucasians	FREM3	FRAS1-related extracellular matrix protein 3. Extracellular matrix protein which may play a role in cell adhesion.	rs7676614	A allele may lead to reduced FREM3 gene expression (a trend in (75))
[^] Nikolova et al., 2015 (75)	N=365 (192 females), age: 19.71	Non Hispanic Caucasians	FREM3		rs1391187 (imputed)	G allele associated with reduced FREM3 gene expression (75)
[^] Arloth et al., 2015 (12)	N=647 (362 females), age: 16.65	Caucasian, African American, Latino, Asian, and other	19 SNPs*	SNPs that are associated with the genetic regulation of GR stimulated gene expression		
Marusak et al., 2015 (110)	N=55 (34 females), age: 11.7	Caucasian, African American, Hispanic, Mixed	OXTR	Oxytocin receptor	rs2254298	Unknown
Liu et al., 2010 (101)	N=68 (29 BD, 42 females), age=14	Mostly Caucasian (6 African American and 5 unknown)	DOK5	Docking protein 5. Plays an important role in neurotrophin signaling pathways, neuronal development, and differentiation.	rs2023454 (used proxy rs16999924)	Unknown
Domschke et al., 2012 (83)	N=85 (51 females) age=37.37	European	COMT	Catechol-O-methyltransferase. Degrades catecholamines such as	rs4680	A (Met) allele alters the structure of COMT and leads to a reduction in its activity (127)

Target Study	Demographics	Ethnicity	Gene	Gene product	Polymorphism	Functionality
Ridder <i>et al.</i> , 2012 (108)	Sample 1: N=60 (22 females), age=21.25; Sample 2: N=52 (20 females), age: 22.27	Missing	NR3C1	dopamine, epinephrine, and norepinephrine Nuclear Receptor Subfamily 3 Group C Member 1. Glucocorticoid receptor. Involved in the negative feedback loop of the HPA axis.	Carriers of 3 or more minor alleles vs all others: rs33389, rs41423247 (used proxy rs853180), rs4986593.	Unknown
Soliman <i>et al.</i> , 2010 (104)	N=72 (33 females), age=25.64	Mixed (Caucasian, Asian, Hispanic, African American)	BDNF	Brain-derived neurotrophic factor. A member of the nerve-growth-factor family. Plays an important role in neuronal survival and development.	Val66Met (rs6265)	Met allele leads to impairments in intracellular trafficking and activity-dependent secretion (128, 129).
Weber <i>et al.</i> , 2016 (109)	N=48 (Panic disorder, 33 females), age=36.06	Caucasian	CRHR1	Corticotropin-releasing hormone/factor receptor 1. Involved in the initiation of the stress response.	rs17689966 (imputed)	G allele associated with reduced mRNA expression in the amygdala and forebrain (109)

^ Study is based on a subsample of our replication sample. ^^ In the original study a replication was attempted with a subsample of our replication sample that included 99 participants. The replication was not successful. * rs11588837, rs8106959, rs10002500, rs1894823, rs2281677, rs12611262, rs10229363, rs9858280, rs4838884, rs2072443, rs2269799 (used proxy rs2285625), rs2395891, rs2422008, rs2956993, rs7534993, rs12901022, rs921320, rs7194275, rs7252014.

Table S3. Studies that were excluded from the replication analyses.

Study	Sample	Ethnicity	Type of scanner	Gene	Gene product description	Polymorphism functionality	Higher activation	Contrast
Numerous studies				SLC6A4 (5-HTTLPR, insertion/deletion)	Solute carrier family 6 member 4. Serotonin transporter	Short allele is associated with reduced transcription and expression of the transporter (Lesch <i>et al.</i> , 1996)	Inconsistent	
Lee & Ham, 2008	N=54 (all females), mean age: 23.2	Asian	1.5T	HTR2A (rs6311, A-1438G)	Serotonin receptor 2A	G allele leads to higher levels of methylation and higher levels of expression (Polesskaya <i>et al.</i> , 2005)	T allele	Sad > neutral (not in angry > neutral)
Lee <i>et al.</i> , 2009	N=26 (all females), mean age=50.3	Asian	1.5T	TPH1 (rs1800532, A218C)	lisoenzyme of tryptophan hydroxylase, the rate-limiting enzyme in the biosynthesis of serotonin	Unknown	A allele	sad > neutral, but not in angry > neutral.
Meyer-Lindenberg <i>et al.</i> , 2006	N=142 (72 females), mean age: 29.64	Caucasians	3T	MAOA (VNTR)	Monoamine oxidase A. Oxidative deamination of amins, serotonin, dopamine, norepinephrine	3.5 and 4 repeats are associated with higher transcription levels (Sabol <i>et al.</i> , 1996)	Low expression variants (2,3, or 5 repeats)	angry and fearful > shapes
Yang <i>et al.</i> 2016	N=18 pathological gamblers (5 females) and 25 controls (10 females), mean age: 32.76	European American	3T	DBH (rs1611115, C-1021T)	Dopamine beta-hydroxylase. Mostly converts dopamine to norepinephrine	T allele predicts low DBH plasma activity (Zabetian <i>et al.</i> , 2001)	CC genotype	Higher activation to videos > the baseline before and after viewing the scenarios
Bergman <i>et al.</i> , 2014	N=85 (44 females), mean age: 45.2	Caucasians	3T	DAT1/SLC6A3 (VNTR)	Dopamine active transporter 1	Mixed. 9-repeat associated with higher levels of DAT in the striatum (van Dyck <i>et al.</i> , 2005; van de Giessen <i>et al.</i> , 2009). Conversely, 9-repeat was associated with lower expression in the putamen (Heinz <i>et al.</i> , 2000) and lower expression in vitro (Fuke <i>et al.</i> , 2001; VanNess <i>et al.</i> , 2005). However, another study did not find a difference between the variants in vitro (Mill <i>et al.</i> , 2005).	9-repeat	angry and fearful > shapes

Study	Sample	Ethnicity	Type of scanner	Gene	Gene product description	Polymorphism functionality	Higher activation	Contrast
Meyer-Lindenberg <i>et al.</i> , 2009	N=121 other details not provided for fMRI subsample	European Caucasians	3T	AVPR1A (RS3)	Vasopressin receptor 1A	Long alleles (>325) associated with higher mRNA in the hippocampus (Knafo <i>et al.</i> , 2008) and shorter alleles have been associated with decreased promoter activity (Tansey <i>et al.</i> , 2011)	334 allele	angry and fearful > shapes
Haram <i>et al.</i> , 2016	N=104 schizophrenia patients (41 females), 100 affective disorder patients (57 females), and 142 controls (58 females). Only significant in SCZ. Mean age:31.5	Caucasians	1.5T	OXTR (rs237902)	Oxytocin receptor	Unknown	AA genotype	angry + fearful > shapes
Jorde <i>et al.</i> , 2014	N=81 abstinent, alcohol-dependent patients (24 females). Mean age: 45.59	Missing (German sample)	3T	GATA4 (rs13273672)	GATA binding protein 4. A transcription factor regulating the transcription of the atrial natriuretic peptide (ANP). Binding of ANP to natriuretic peptide receptors has been found to inhibit noradrenergic and dopaminergic neurotransmission. In addition, natriuretic peptides reduce the release of corticotrophin-releasing hormone (CRH) receptors	Reduced variability of ANP in carriers of at least one G allele. (Keifer <i>et al.</i> , 2011)	AA genotype	alcohol > neutral cues
Kuhn <i>et al.</i> , 2016	N=49 (25 females), mean age: 25		3T	NOS1 (exon 1f - VNTR)	Nitric oxide synthase 1	Long alleles are associated with higher gene expression levels (Kuhn <i>et al.</i> , 2016)	Short allele	unpredictable context > safe context

Study	Sample	Ethnicity	Type of scanner	Gene	Gene product description	Polymorphism functionality	Higher activation	Contrast
Heinrich et al., 2013	N=485 (248 females), mean age: 14.26	Missing	3T	ODZ4 (rs12576775)	The protein encoded by this gene plays a role in establishing proper neuronal connectivity during development.	unknown	G allele	No effect on face task. Higher activation during reward sensitivity and reward expectation.
Rasch et al. 2009;	N=57 (41 females), mean age: 24.1	Caucasians	3T	ADRA2B (deletion)	Presynaptic noradrenergic α 2B receptor	Deletion shows both agonistic and antagonistic effects (Small et al., 2001), and needs to be clarified	deletion	negative > neutral and positive > neutral
Stevens et al., 2014	N=49 PTSD risk females, mean age: 38.7.	African American	3T	ADCYAP1R1 (rs2267735)	Adenylate cyclase activating polypeptide 1 receptor type 1. The pituitary adenylate cyclase-activating polypeptide (PACAP) receptor.	Females with the CC genotype express significantly less ADCYAP1R1 mRNA than males, or than females who are G carriers. (Ressler et al., 2011)	CC genotype	fearful > neutral
Hessl et al., 2007	N=21 males (11 with the FMR1 premutation), mean age: 42.9	Caucasians, Hispanic, East Indian	1.5T	FMR1 (CGG repeat)	Fragile X mental retardation 1. Codes for fragile X mental retardation protein which is involved in RNA binding and translation of proteins. May be involved in synaptic plasticity.	higher repeats in premutation alleles associated with higher mRNA levels (Tassone et al., 2000)		fearful > scrambled (lower activity in premutation carriers) neutral > scrambled (higher activity in premutation carriers)
Whalley et al., 2012	N=73 bipolar disorder risk and 52 controls. No details are given for the fMRI subsample	Missing	1.5T	A polygenic risk score for BD based on GWAS				sentence completion > baseline

Table S4. Hardy Weinberg equilibrium of all included SNPs

CHR	SNP	A1	A2	HWE original p values				
				Caucasians	Latino	African American	Asians 1	Asians 2
1	rs324420	A	C	0.8913	1	0.4122	0.6796	0.808
1	rs4838884	G	A	0.7754	1	1	0.4984	1
1	rs11588837	G	A	1	1	1	1	1
1	rs3767488	G	A	0.4243	1	0.3615	0.7615	0.5721
1	rs7534993	A	G	0.702	0.6117	0.2098	0.44	0.3857
1	rs10799590	A	G	0.2873	0.2596	0.6457	0.6308	1
2	rs921320	A	C	0.1928	0.6483	0.7865	1	0.7696
2	rs2422008	A	C	0.6602	0.1106	0.03496	0.6258	1
2	rs12620091	C	T	0.8519	1	0.03139	0.2965	1
2	rs333229	T	G	0.4853	0.1212	0.09176	1	0.7981
2	rs16944	A	G	0.9225	0.4816	0.4747	0.6308	0.5027
2	rs860573	A	G	N/A	N/A	N/A	0.07888	1
3	rs1042778	T	G	1	0.8114	0.855	0.2244	1
3	rs2254298	A	G	0.03175	0.6899	0.01863	0.3463	0.8781
3	rs53576	A	G	0.7645	0.5613	0.7967	0.6076	0.7543
3	rs9858280	C	T	0.7067	0.7879	1	0.7289	1
4	rs10014254	T	C	1	1	1	1	1
4	rs10002500	T	C	0.6689	0.6811	0.2192	0.3527	0.649
4	rs1391187	A	G	0.1147	0.253	0.3678	0.05897	0.5447
4	rs7676614	G	A	0.1893	1	1	0.7963	1
4	rs5522	C	T	0.3768	1	1	1	1
5	rs358527	T	C	0.6003	0.4909	0.08443	0.8105	1
5	rs853180	G	A	0.05168	0.2491	0.5924	0.03044	0.005043
5	rs4986593	G	A	0.7165	1	1	0.04808	0.04186
5	rs33389	T	C	0.5842	0.1056	0.3648	1	0.2045
6	rs1360780	T	C	0.9188	0.608	1	1	0.0691

CHR	SNP	A1	A2	HWE original p values				
				Caucasians	Latino	African American	Asians 1	Asians 2
7	rs10254767	G	A	0.3065	0.4979	1	0.7835	1
7	rs17149106	T	G	1	1	1	0.3507	1
7	rs16147	T	C	0.1617	0.2497	0.5775	0.3338	1
7	rs16139	C	T	1	1	1	0.2396	1
7	rs5573	G	A	0.1896	0.2357	0.573	0.4687	1
7	rs5574	T	C	0.2573	0.02153	0.7007	0.7609	0.4722
7	rs324987	C	T	0.9303	0.05445	1	0.2375	0.5891
7	rs10229363	G	A	0.4091	1	0.4282	1	1
7	rs13227433	G	T	0.521	0.7638	1	0.53	0.6101
7	rs2522833	C	A	0.3275	0.82	1	1	0.4433
7	rs2072443	T	C	0.9286	1	0.5647	0.3467	0.4214
8	rs1390938	A	G	0.3693	0.1755	0.5998	0.5968	0.5263
11	rs6265	T	C	0.8919	1	1	0.1052	0.1801
11	rs2956993	G	T	0.221	0.4472	0.2394	0.4942	1
11	rs795467	A	G	0.2624	0.5988	1	1	0.7467
11	rs1946518	T	G	0.592	0.03973	0.5709	1	0.8934
11	rs1800497	A	G	0.4092	0.3347	0.8399	0.2339	0.2153
11	rs1076560	A	C	0.5042	0.4491	0.09248	0.3602	0.3393
11	rs1062613	T	C	0.897	1	0.376	0.3527	1
12	rs1006737	A	G	0.229	0.8042	1	1	0.3596
12	rs1894823	T	C	0.05853	0.2041	0.614	1	0.109
12	rs10875995	C	T	0.6934	1	0.03916	0.7969	1
12	rs1861494	C	T	0.06035	0.5184	1	0.6177	0.3084
12	rs4570625	T	G	0.152	0.7512	1	0.6051	0.2166
13	rs9537793	G	A	0.4816	1	0.3768	0.3312	0.7853
14	rs2281677	A	G	0.9262	0.6117	0.7222	0.6341	0.2728
15	rs12901022	T	C	0.4873	0.361	0.0825	0.6276	0.7477
15	rs2285625	C	T	0.1323	1	1	0.2742	0.595

CHR	SNP	A1	A2	HWE original p values				
				Caucasians	Latino	African American	Asians 1	Asians 2
16	rs7194275	C	T	0.4723	1	0.4754	1	0.1647
16	rs1064448	G	T	0.9305	0.4929	0.7816	0.4238	1
17	rs17689966	G	A	0.9274	1	1	0.3826	0.2682
19	rs2395891	T	G	0.334	0.7874	0.1497	0.007006	1
19	rs12611262	T	C	0.168	0.8147	0.3212	0.3376	0.5516
19	rs7252014	A	G	0.4345	1	0.2968	0.6308	0.25
19	rs8106959	G	A	0.2571	1	0.614	0.2791	1
20	rs16999924	A	G	0.3832	1	1	1	1
22	rs4680	G	A	0.09833	0.1714	1	1	0.4995

Note. All p values were higher than the lowest FDR corrected p value of ~ 0.00016 . CHR=chromosome number.

Supplemental References

1. Loth E, Poline J-B, Thyreau B, Jia T, Tao C, Lourdasamy A, et al. (2014): Oxytocin receptor genotype modulates ventral striatal activity to social cues and response to stressful life events. *Biol Psychiatry*. 76:367-376.
2. Lecrubier Y, Sheehan DV, Weiller E, Amorim P, Bonora I, Sheehan KH, et al. (1997): The Mini International Neuropsychiatric Interview (MINI). A short diagnostic structured interview: reliability and validity according to the CIDI. *Eur Psychiatry*. 12:224-231.
3. First MB, Spitzer RL, Gibbon M, Williams JBM (1996): *Structured Clinical Interview for DSM-IV Axis I Disorders, Research Version, Nonpatient Edition*. New York: New York State Psychiatric Institute, Biometrics Research Department.
4. Howie B, Marchini J, Stephens M (2011): Genotype imputation with thousands of genomes. *G3: Genes, Genomes, Genetics*. 1:457-470.
5. Delaneau O, Marchini J, Zagury J-F (2012): A linear complexity phasing method for thousands of genomes. *Nat Methods*. 9:179-181.
6. Klein A, Andersson J, Ardekani BA, Ashburner J, Avants B, Chiang M-C, et al. (2009): Evaluation of 14 nonlinear deformation algorithms applied to human brain MRI registration. *Neuroimage*. 46:786-802.
7. Cox RW (1996): AFNI: software for analysis and visualization of functional magnetic resonance neuroimages. *Comput Biomed Res*. 29:162-173.
8. Greve DN, Fischl B (2009): Accurate and robust brain image alignment using boundary-based registration. *Neuroimage*. 48:63-72.
9. Nichols TE (2017): Notes on creating a standardized version of DVARS. *arXiv preprint arXiv:170401469*.
10. Power JD, Mitra A, Laumann TO, Snyder AZ, Schlaggar BL, Petersen SE (2014): Methods to detect, characterize, and remove motion artifact in resting state fMRI. *Neuroimage*. 84:320-341.
11. Tyszka JM, Pauli WM (2016): In vivo delineation of subdivisions of the human amygdaloid complex in a high-resolution group template. *Hum Brain Mapp*. 37:3979-3998.
12. Arloth J, Bogdan R, Weber P, Frishman G, Menke A, Wagner KV, et al. (2015): Genetic differences in the immediate transcriptome response to stress predict risk-related brain function and psychiatric disorders. *Neuron*. 86:1189-1202.
13. Swartz JR, Prather AA, Di Iorio CR, Bogdan R, Hariri AR (2016): A Functional Interleukin-18 Haplotype Predicts Depression and Anxiety through Increased Threat-Related Amygdala Reactivity in Women but Not Men. *Neuropsychopharmacology*.
14. Oathes DJ, Hilt LM, Nitschke JB (2015): Affective Neural Responses Modulated by Serotonin Transporter Genotype in Clinical Anxiety and Depression. *PLoS ONE*. 10:e0115820.
15. Schneck N, Miller JM, Delorenzo C, Kikuchi T, Sublette ME, Oquendo MA, et al. (2016): Relationship of the serotonin transporter gene promoter polymorphism (5-HTTLPR)

- genotype and serotonin transporter binding to neural processing of negative emotional stimuli. *J Affect Disord.* 190:494-498.
16. Stoeckel MC, Esser RW, Gamer M, Kalisch R, Büchel C, von Leupoldt A (2015): Amygdala response to anticipation of dyspnea is modulated by 5-HTTLPR genotype. *Psychophysiology.* 52:973-976.
 17. Bastiaansen JA, Servaas MN, Marsman JBC, Ormel J, Nolte IM, Riese H, et al. (2014): Filling the gap: relationship between the serotonin-transporter-linked polymorphic region and amygdala activation. *Psychol Sci.* 25:2058-2066.
 18. Klumpp H, Fitzgerald DA, Cook E, Shankman SA, Angstadt M, Phan KL (2014): Serotonin transporter gene alters insula activity to threat in social anxiety disorder. *Neuroreport.* 25:926-931.
 19. Costafreda SG, McCann P, Saker P, Cole JH, Cohen-Woods S, Farmer AE, et al. (2013): Modulation of amygdala response and connectivity in depression by serotonin transporter polymorphism and diagnosis. *J Affect Disord.* 150:96-103.
 20. Klucken T, Wehrum S, Schweckendiek J, Merz CJ, Hennig J, Vaitl D, et al. (2013): The 5-HTTLPR polymorphism is associated with altered hemodynamic responses during appetitive conditioning. *Hum Brain Mapp.* 34:2549-2560.
 21. O'Nions EJ, Dolan RJ, Roiser JP (2011): Serotonin transporter genotype modulates subgenual response to fearful faces using an incidental task. *J Cogn Neurosci.* 23:3681-3693.
 22. Morey RA, Hariri AR, Gold AL, Hauser MA, Munger HJ, Dolcos F, et al. (2011): Serotonin transporter gene polymorphisms and brain function during emotional distraction from cognitive processing in posttraumatic stress disorder. *BMC psychiatry.* 11:76.
 23. Lonsdorf TB, Golkar A, Lindström KM, Fransson P, Schalling M, Öhman A, et al. (2011): 5-HTTLPR and COMTval158met genotype gate amygdala reactivity and habituation. *Biol Psychol.* 87:106-112.
 24. Lemogne C, Gorwood P, Boni C, Pessiglione M, Lehericy S, Fossati P (2011): Cognitive appraisal and life stress moderate the effects of the 5-HTTLPR polymorphism on amygdala reactivity. *Hum Brain Mapp.* 32:1856-1867.
 25. Kobiella A, Reimold M, Ulshöfer D, Ikonomidou V, Vollmert C, Vollstädt-Klein S, et al. (2011): How the serotonin transporter 5-HTTLPR polymorphism influences amygdala function: the roles of in vivo serotonin transporter expression and amygdala structure. *Translational Psychiatry.* 1:e37.
 26. Furman DJ, Hamilton JP, Joormann J, Gotlib IH (2010): Altered timing of amygdala activation during sad mood elaboration as a function of 5-HTTLPR. *Soc Cogn Affect Neurosci.* 6:270-276.
 27. Dannlowski U, Kugel H, Franke F, Stuhrmann A, Hohoff C, Zwanzger P, et al. (2011): Neuropeptide-S (NPS) receptor genotype modulates basolateral amygdala responsiveness to aversive stimuli. *Neuropsychopharmacology.* 36:1879-1885.
 28. Gillihan SJ, Rao H, Brennan L, Wang DJ, Detre JA, Sankoorikal GMV, et al. (2011): Serotonin transporter genotype modulates the association between depressive

- symptoms and amygdala activity among psychiatrically healthy adults. *Psychiatry Research: Neuroimaging*. 193:161-167.
29. Lau JY, Goldman D, Buzas B, Fromm SJ, Guyer AE, Hodgkinson C, et al. (2009): Amygdala function and 5-HTT gene variants in adolescent anxiety and major depressive disorder. *Biol Psychiatry*. 65:349-355.
 30. Rao H, Gillihan SJ, Wang J, Korczykowski M, Sankoorikal GMV, Kaercher KA, et al. (2007): Genetic variation in serotonin transporter alters resting brain function in healthy individuals. *Biol Psychiatry*. 62:600-606.
 31. Dannlowski U, Ohrmann P, Bauer J, Deckert J, Hohoff C, Kugel H, et al. (2008): 5-HTTLPR biases amygdala activity in response to masked facial expressions in major depression. *Neuropsychopharmacology*. 33:418-424.
 32. Heinz A, Smolka MN, Braus DF, Wrase J, Beck A, Flor H, et al. (2007): Serotonin transporter genotype (5-HTTLPR): effects of neutral and undefined conditions on amygdala activation. *Biol Psychiatry*. 61:1011-1014.
 33. Bertolino A, Arciero G, Rubino V, Latorre V, De Candia M, Mazzola V, et al. (2005): Variation of human amygdala response during threatening stimuli as a function of 5' HTTLPR genotype and personality style. *Biol Psychiatry*. 57:1517-1525.
 34. Hariri AR, Drabant EM, Munoz KE, Kolachana BS, Mattay VS, Egan MF, et al. (2005): A susceptibility gene for affective disorders and the response of the human amygdala. *Arch Gen Psychiatry*. 62:146-152.
 35. Hariri AR, Mattay VS, Tessitore A, Kolachana B, Fera F, Goldman D, et al. (2002): Serotonin transporter genetic variation and the response of the human amygdala. *Science*. 297:400-403.
 36. Lee BT, Ham BJ (2008): Serotonergic genes and amygdala activity in response to negative affective facial stimuli in Korean women. *Genes Brain Behav*. 7:899-905.
 37. Smolka M, Bühler M, Schumann G, Klein S, Hu X, Moayer M, et al. (2007): Gene–gene effects on central processing of aversive stimuli. *Mol Psychiatry*. 12:307-317.
 38. Lee B-T, Ham B-J (2008): Monoamine oxidase A–uVNTR genotype affects limbic brain activity in response to affective facial stimuli. *Neuroreport*. 19:515-519.
 39. Kuhn M, Haaker J, Glotzbach-Schoon E, Schümann D, Andreatta M, Mechias M-L, et al. (2016): Converging evidence for an impact of a functional NOS gene variation on anxiety-related processes. *Social cognitive and affective neuroscience*. 11:803-812.
 40. Meyer-Lindenberg A, Buckholtz JW, Kolachana B, Hariri AR, Pezawas L, Blasi G, et al. (2006): Neural mechanisms of genetic risk for impulsivity and violence in humans. *Proc Natl Acad Sci U S A*. 103:6269-6274.
 41. Denson TF, Dobson-Stone C, Ronay R, von Hippel W, Schira MM (2014): A functional polymorphism of the MAOA gene is associated with neural responses to induced anger control. *J Cogn Neurosci*. 26:1418-1427.
 42. Wetherill RR, Jagannathan K, Lohoff FW, Ehrman R, O'brien CP, Childress AR, et al. (2014): Neural correlates of attentional bias for smoking cues: modulation by variance in the dopamine transporter gene. *Addict Biol*. 19:294-304.

43. Bergman O, Åhs F, Furmark T, Appel L, Linnman C, Faria V, et al. (2014): Association between amygdala reactivity and a dopamine transporter gene polymorphism. *Translational Psychiatry*. 4:e420.
44. Manuck SB, Marsland AL, Flory JD, Gorka A, Ferrell RE, Hariri AR (2010): Salivary testosterone and a trinucleotide (CAG) length polymorphism in the androgen receptor gene predict amygdala reactivity in men. *Psychoneuroendocrinology*. 35:94-104.
45. Rasch B, Spalek K, Buholzer S, Luechinger R, Boesiger P, Papassotiropoulos A, et al. (2009): A genetic variation of the noradrenergic system is related to differential amygdala activation during encoding of emotional memories. *Proc Natl Acad Sci U S A*. 106:19191-19196.
46. Cousijn H, Rijpkema M, Qin S, van Marle HJ, Franke B, Hermans EJ, et al. (2010): Acute stress modulates genotype effects on amygdala processing in humans. *Proc Natl Acad Sci U S A*. 107:9867-9872.
47. Urner M, van Wingen G, Franke B, Rijpkema M, Fernández G, Tendolkar I (2011): Genetic variation of the α 2b-adrenoceptor affects neural correlates of successful emotional memory formation. *Hum Brain Mapp*. 32:2096-2103.
48. Li S, Weerda R, Milde C, Wolf OT, Thiel CM (2015): ADRA2B genotype differentially modulates stress-induced neural activity in the amygdala and hippocampus during emotional memory retrieval. *Psychopharmacology (Berl)*. 232:755.
49. Meyer-Lindenberg A, Kolachana B, Gold B, Olsh A, Nicodemus K, Mattay V, et al. (2009): Genetic variants in AVPR1A linked to autism predict amygdala activation and personality traits in healthy humans. *Mol Psychiatry*. 14:968-975.
50. Hessel D, Rivera S, Koldewyn K, Cordeiro L, Adams J, Tassone F, et al. (2007): Amygdala dysfunction in men with the fragile X premutation. *Brain*. 130:404-416.
51. Whalley H, Papmeyer M, Sprooten E, Romaniuk L, Blackwood D, Glahn D, et al. (2012): The influence of polygenic risk for bipolar disorder on neural activation assessed using fMRI. *Translational Psychiatry*. 2:e130.
52. Heinrich A, Lourdasamy A, Tzschoppe J, Vollstädt-Klein S, Bühler M, Steiner S, et al. (2013): The risk variant in ODZ4 for bipolar disorder impacts on amygdala activation during reward processing. *Bipolar Disord*. 15:440-445.
53. Jorde A, Bach P, Witt SH, Becker K, Reinhard I, Vollstädt-Klein S, et al. (2014): Genetic variation in the atrial natriuretic peptide transcription factor GATA4 modulates amygdala responsiveness in alcohol dependence. *Biol Psychiatry*. 75:790-797.
54. Lee BT, Lee HY, Lee BC, Pae CU, Yoon BJ, Ryu SG, et al. (2009): Impact of the tryptophan hydroxylase 1 gene A218C polymorphism on amygdala activity in response to affective facial stimuli in patients with major depressive disorder. *Genes Brain Behav*. 8:512-518.
55. Yang B-Z, Balodis IM, Lacadie CM, Xu J, Potenza MN (2016): A preliminary study of DBH (Encoding Dopamine Beta-Hydroxylase) genetic variation and neural correlates of emotional and motivational processing in individuals with and without pathological gambling. *Journal of behavioral addictions*. 5:282-292.

56. Haram M, Bettella F, Brandt CL, Quintana DS, Nerhus M, Bjella T, et al. (2016): Contribution of oxytocin receptor polymorphisms to amygdala activation in schizophrenia spectrum disorders. *British Journal of Psychiatry Open*. 2:353-358.
57. Stevens JS, Almli LM, Fani N, Gutman DA, Bradley B, Norrholm SD, et al. (2014): PACAP receptor gene polymorphism impacts fear responses in the amygdala and hippocampus. *Proc Natl Acad Sci U S A*. 111:3158-3163.
58. Chang H, Hoshina N, Zhang C, Ma Y, Cao H, Wang Y, et al. (2017): The protocadherin 17 gene affects cognition, personality, amygdala structure and function, synapse development and risk of major mood disorders. *Mol Psychiatry*.
59. Fakra E, Hyde LW, Gorka A, Fisher PM, Muñoz KE, Kimak M, et al. (2009): Effects of HTR1A C (- 1019) G on amygdala reactivity and trait anxiety. *Arch Gen Psychiatry*. 66:33-40.
60. Domschke K, Braun M, Ohrmann P, Suslow T, Kugel H, Bauer J, et al. (2006): Association of the functional- 1019C/G 5-HT 1A polymorphism with prefrontal cortex and amygdala activation measured with 3 T fMRI in panic disorder. *The The International Journal of Neuropsychopharmacology*. 9:349-355.
61. Straube B, Reif A, Richter J, Lueken U, Weber H, Arolt V, et al. (2014): The functional- 1019C/G HTR1A polymorphism and mechanisms of fear. *Translational Psychiatry*. 4:e490.
62. Smoller JW, Paulus MP, Fagerness JA, Purcell S, Yamaki LH, Hirshfeld-Becker D, et al. (2008): Influence of RGS2 on anxiety-related temperament, personality, and brain function. *Arch Gen Psychiatry*. 65:298-308.
63. Smoller JW, Gallagher PJ, Duncan LE, McGrath LM, Haddad SA, Holmes AJ, et al. (2014): The human ortholog of acid-sensing ion channel gene ASIC1a is associated with panic disorder and amygdala structure and function. *Biol Psychiatry*. 76:902-910.
64. Hariri AR, Gorka A, Hyde LW, Kimak M, Halder I, Ducci F, et al. (2009): Divergent effects of genetic variation in endocannabinoid signaling on human threat-and reward-related brain function. *Biol Psychiatry*. 66:9-16.
65. Gunduz-Cinar O, MacPherson K, Cinar R, Gamble-George J, Sugden K, Williams B, et al. (2013): Convergent translational evidence of a role for anandamide in amygdala-mediated fear extinction, threat processing and stress-reactivity. *Mol Psychiatry*. 18:813-823.
66. Bogdan R, Williamson DE, Hariri AR (2012): Mineralocorticoid receptor Iso/Val (rs5522) genotype moderates the association between previous childhood emotional neglect and amygdala reactivity. *Am J Psychiatry*. 169:515-522.
67. Wellman CL, Camp M, Jones VM, MacPherson KP, Ihne J, Fitzgerald P, et al. (2013): Convergent effects of mouse Pet-1 deletion and human PET-1 variation on amygdala fear and threat processing. *Exp Neurol*. 250:260-269.
68. Neumann SA, Brown SM, Ferrell RE, Flory JD, Manuck SB, Hariri AR (2006): Human choline transporter gene variation is associated with corticolimbic reactivity and autonomic-cholinergic function. *Biol Psychiatry*. 60:1155-1162.

69. Holz NE, Buchmann AF, Boecker R, Blomeyer D, Baumeister S, Wolf I, et al. (2015): Role of FKBP5 in emotion processing: results on amygdala activity, connectivity and volume. *Brain Struct Funct.* 220:1355-1368.
70. Tost H, Kolachana B, Hakimi S, Lemaitre H, Verchinski BA, Mattay VS, et al. (2010): A common allele in the oxytocin receptor gene (OXTR) impacts prosocial temperament and human hypothalamic-limbic structure and function. *Proc Natl Acad Sci U S A.* 107:13936-13941.
71. Redlich R, Stacey D, Opel N, Grotegerd D, Dohm K, Kugel H, et al. (2015): Evidence of an IFN- γ by early life stress interaction in the regulation of amygdala reactivity to emotional stimuli. *Psychoneuroendocrinology.* 62:166-173.
72. Joeyen-Waldorf J, Nikolova YS, Edgar N, Walsh C, Kota R, Lewis DA, et al. (2012): Adenylate cyclase 7 is implicated in the biology of depression and modulation of affective neural circuitry. *Biol Psychiatry.* 71:627-632.
73. Swartz JR, Waller R, Bogdan R, Knodt AR, Sabhlok A, Hyde LW, et al. (2015): A common polymorphism in a Williams syndrome gene predicts amygdala reactivity and extraversion in healthy adults. *Biol Psychiatry.*
74. Lohoff FW, Hodge R, Narasimhan S, Nall A, Ferraro TN, Mickey BJ, et al. (2014): Functional genetic variants in the vesicular monoamine transporter 1 modulate emotion processing. *Mol Psychiatry.* 19:129-139.
75. Nikolova YS, Iruku SP, Lin C-W, Conley ED, Puralewski R, French B, et al. (2015): FRAS1-related extracellular matrix 3 (FREM3) single-nucleotide polymorphism effects on gene expression, amygdala reactivity and perceptual processing speed: An accelerated aging pathway of depression risk. *Frontiers in psychology.* 6.
76. Tesli M, Skatun KC, Ousdal OT, Brown AA, Thoresen C, Agartz I, et al. (2013): CACNA1C risk variant and amygdala activity in bipolar disorder, schizophrenia and healthy controls. *PLoS ONE.* 8:e56970.
77. Sumner JA, Sheridan MA, Drury SS, Esteves KC, Walsh K, Koenen KC, et al. (2015): Variation in CACNA1C is associated with amygdala structure and function in adolescents. *J Child Adolesc Psychopharmacol.* 25:701-710.
78. Jogia J, Ruberto G, Lelli-Chiesa G, Vassos E, Maieru M, Tatarelli R, et al. (2011): The impact of the CACNA1C gene polymorphism on frontolimbic function in bipolar disorder. *Molecular psychiatry.* 16.
79. Wessa M, Linke J, Witt S, Nieratschker V, Esslinger C, Kirsch P, et al. (2010): The CACNA1C risk variant for bipolar disorder influences limbic activity. *Mol Psychiatry.* 15:1126-1128.
80. Ousdal OT, Brown AA, Jensen J, Nakstad PH, Melle I, Agartz I, et al. (2012): Associations between variants near a monoaminergic pathways gene (PHOX2B) and amygdala reactivity: a genome-wide functional imaging study. *Twin Res Hum Genet.* 15:273-285.
81. Waller R, Corral-Frías NS, Vannucci B, Bogdan R, Knodt AR, Hariri AR, et al. (2016): An oxytocin receptor polymorphism predicts amygdala reactivity and antisocial behavior in men. *Soc Cogn Affect Neurosci.*nsw042.

82. Nelson EC, Agrawal A, Heath AC, Bogdan R, Sherva R, Zhang B, et al. (2016): Evidence of CNH3 involvement in opioid dependence. *Mol Psychiatry*. 21:608.
83. Domschke K, Baune BT, Havlik L, Stuhmann A, Suslow T, Kugel H, et al. (2012): Catechol-O-methyltransferase gene variation: impact on amygdala response to aversive stimuli. *Neuroimage*. 60:2222-2229.
84. Kempton MJ, Haldane M, Jogia J, Christodoulou T, Powell J, Collier D, et al. (2009): The effects of gender and COMT Val158Met polymorphism on fearful facial affect recognition: a fMRI study. *Int J Neuropsychopharmacol*. 12:371-381.
85. Smolka MN, Schumann G, Wrase J, Grüsser SM, Flor H, Mann K, et al. (2005): Catechol-O-methyltransferase val158met genotype affects processing of emotional stimuli in the amygdala and prefrontal cortex. *J Neurosci*. 25:836-842.
86. Drabant EM, Hariri AR, Meyer-Lindenberg A, Munoz KE, Mattay VS, Kolachana BS, et al. (2006): Catechol O-methyltransferase val158met genotype and neural mechanisms related to affective arousal and regulation. *Arch Gen Psychiatry*. 63:1396-1406.
87. Klucken T, Kruse O, Wehrum-Osinsky S, Hennig J, Schweckendiek J, Stark R (2015): Impact of COMT Val158Met-polymorphism on appetitive conditioning and amygdala/prefrontal effective connectivity. *Hum Brain Mapp*. 36:1093-1101.
88. Lelli-Chiesa G, Kempton M, Jogia J, Tatarelli R, Girardi P, Powell J, et al. (2011): The impact of the Val158Met catechol-O-methyltransferase genotype on neural correlates of sad facial affect processing in patients with bipolar disorder and their relatives. *Psychological medicine*. 41:779-788.
89. Rasch B, Spalek K, Buholzer S, Luechinger R, Boesiger P, De Quervain D-F, et al. (2010): Aversive stimuli lead to differential amygdala activation and connectivity patterns depending on catechol-O-methyltransferase Val158Met genotype. *Neuroimage*. 52:1712-1719.
90. Domschke K, Ohrmann P, Braun M, Suslow T, Bauer J, Hohoff C, et al. (2008): Influence of the catechol-O-methyltransferase val158met genotype on amygdala and prefrontal cortex emotional processing in panic disorder. *Psychiatry Research: Neuroimaging*. 163:13-20.
91. Opmeer EM, Kortekaas R, van Tol M-J, van der Wee NJ, Woudstra S, van Buchem MA, et al. (2013): Influence of COMT val158met genotype on the depressed brain during emotional processing and working memory. *PLoS ONE*. 8:e73290.
92. Williams LM, Gatt JM, Grieve SM, Dobson-Stone C, Paul RH, Gordon E, et al. (2010): COMT Val 108/158 Met polymorphism effects on emotional brain function and negativity bias. *Neuroimage*. 53:918-925.
93. Kilpatrick LA, Labus JS, Coveleskie K, Hammer C, Rappold G, Tillisch K, et al. (2011): The HTR3A polymorphism c.-42C> T is associated with amygdala responsiveness in patients with irritable bowel syndrome. *Gastroenterology*. 140:1943-1951.
94. Iidaka T, Ozaki N, Matsumoto A, Nogawa J, Kinoshita Y, Suzuki T, et al. (2005): A variant C178T in the regulatory region of the serotonin receptor gene HTR3A modulates neural activation in the human amygdala. *J Neurosci*. 25:6460-6466.

95. Lee B-T, Lee H-Y, Han C, Pae C-U, Tae WS, Lee M-S, et al. (2011): DRD2/ANKK1 TaqI A polymorphism affects corticostriatal activity in response to negative affective facial stimuli. *Behav Brain Res.* 223:36-41.
96. Canli T, Congdon E, Gutknecht L, Constable R, Lesch K (2005): Amygdala responsiveness is modulated by tryptophan hydroxylase-2 gene variation. *J Neural Transm.* 112:1479-1485.
97. Brown S, Peet E, Manuck S, Williamson D, Dahl R, Ferrell R, et al. (2005): A regulatory variant of the human tryptophan hydroxylase-2 gene biases amygdala reactivity. *Mol Psychiatry.* 10:884-888.
98. Blasi G, Bianco LL, Taurisano P, Gelao B, Romano R, Fazio L, et al. (2009): Functional variation of the dopamine D2 receptor gene is associated with emotional control as well as brain activity and connectivity during emotion processing in humans. *J Neurosci.* 29:14812-14819.
99. Woudstra S, Bochdanovits Z, Van Tol M, Veltman D, Zitman F, Van Buchem M, et al. (2012): Piccolo genotype modulates neural correlates of emotion processing but not executive functioning. *Translational Psychiatry.* 2:e99.
100. Baune BT, Dannlowski U, Domschke K, Janssen DG, Jordan MA, Ohrmann P, et al. (2010): The interleukin 1 beta (IL1B) gene is associated with failure to achieve remission and impaired emotion processing in major depression. *Biol Psychiatry.* 67:543-549.
101. Liu X, Akula N, Skup M, Brotman MA, Leibenluft E, McMahon FJ (2010): A genome-wide association study of amygdala activation in youths with and without bipolar disorder. *J Am Acad Child Adolesc Psychiatry.* 49:33-41.
102. Zhou Z, Zhu G, Hariri AR, Enoch M-A, Scott D, Sinha R, et al. (2008): Genetic variation in human NPY expression affects stress response and emotion. *Nature.* 452:997-1001.
103. Domschke K, Dannlowski U, Hohoff C, Ohrmann P, Bauer J, Kugel H, et al. (2010): Neuropeptide Y (NPY) gene: impact on emotional processing and treatment response in anxious depression. *Eur Neuropsychopharmacol.* 20:301-309.
104. Soliman F, Glatt CE, Bath KG, Levita L, Jones RM, Pattwell SS, et al. (2010): A genetic variant BDNF polymorphism alters extinction learning in both mouse and human. *Science.* 327:863-866.
105. Montag C, Reuter M, Newport B, Elger C, Weber B (2008): The BDNF Val66Met polymorphism affects amygdala activity in response to emotional stimuli: evidence from a genetic imaging study. *Neuroimage.* 42:1554-1559.
106. Lau JY, Goldman D, Buzas B, Hodgkinson C, Leibenluft E, Nelson E, et al. (2010): BDNF gene polymorphism (Val66Met) predicts amygdala and anterior hippocampus responses to emotional faces in anxious and depressed adolescents. *Neuroimage.* 53:952-961.
107. Gasic G, Smoller J, Perlis R, Sun M, Lee S, Kim B, et al. (2009): BDNF, relative preference, and reward circuitry responses to emotional communication. *American Journal of Medical Genetics Part B: Neuropsychiatric Genetics.* 150:762-781.
108. Ridder S, Treutlein J, Nees F, Lang S, Diener S, Wessa M, et al. (2012): Brain activation during fear conditioning in humans depends on genetic variations related to

- functioning of the hypothalamic–pituitary–adrenal axis: first evidence from two independent subsamples. *Psychol Med.* 42:2325-2335.
109. Weber H, Richter J, Straube B, Lueken U, Domschke K, Schartner C, et al. (2016): Allelic variation in CRHR1 predisposes to panic disorder: evidence for biased fear processing. *Mol Psychiatry.* 21:813.
 110. Marusak HA, Furman DJ, Kuruvadi N, Shattuck DW, Joshi SH, Joshi AA, et al. (2015): Amygdala responses to salient social cues vary with oxytocin receptor genotype in youth. *Neuropsychologia.* 79:1-9.
 111. Fakra E, Hyde LW, Gorka A, Fisher PM, Munoz KE, Kimak M, et al. (2009): Effects of HTR1A C (- 1019) G on amygdala reactivity and trait anxiety. *Arch Gen Psychiatry.* 66:33-40.
 112. Lemonde S, Turecki G, Bakish D, Du L, Hrdina PD, Bown CD, et al. (2003): Impaired repression at a 5-hydroxytryptamine 1A receptor gene polymorphism associated with major depression and suicide. *J Neurosci.* 23:8788-8799.
 113. Bernier V, Stocco R, Bogusky MJ, Joyce JG, Parachoniak C, Grenier K, et al. (2006): Structure/function relationships in the neuropeptide s receptor: molecular consequences of the asthma-associated mutation N107I. *J Biol Chem.*
 114. Reinscheid RK, Xu Y-L, Okamura N, Zeng J, Chung S, Pai R, et al. (2005): Pharmacological characterization of human and murine neuropeptide s receptor variants. *J Pharmacol Exp Ther.* 315:1338-1345.
 115. Semplicini A, Lenzini L, Sartori M, Papparella I, Calò LA, Pagnin E, et al. (2006): Reduced expression of regulator of G-protein signaling 2 (RGS2) in hypertensive patients increases calcium mobilization and ERK1/2 phosphorylation induced by angiotensin II. *J Hypertens.* 24:1115-1124.
 116. DeRijk RH, Wüst S, Meijer OC, Zennaro M-C, Federenko IS, Hellhammer DH, et al. (2006): A common polymorphism in the mineralocorticoid receptor modulates stress responsiveness. *The Journal of Clinical Endocrinology & Metabolism.* 91:5083-5089.
 117. Van Leeuwen N, Kumsta R, Entringer S, de Kloet ER, Zitman FG, DeRijk RH, et al. (2010): Functional mineralocorticoid receptor (MR) gene variation influences the cortisol awakening response after dexamethasone. *Psychoneuroendocrinology.* 35:339-349.
 118. Binder EB, Salyakina D, Lichtner P, Wochnik GM, Ising M, Pütz B, et al. (2004): Polymorphisms in FKBP5 are associated with increased recurrence of depressive episodes and rapid response to antidepressant treatment. *Nat Genet.* 36:1319-1325.
 119. Binder EB (2009): The role of FKBP5, a co-chaperone of the glucocorticoid receptor in the pathogenesis and therapy of affective and anxiety disorders. *Psychoneuroendocrinology.* 34:S186-S195.
 120. Gonsky R, Deem RL, Landers CJ, Haritunians T, Yang S, Targan SR (2014): IFNG rs1861494 polymorphism is associated with IBD disease severity and functional changes in both IFNG methylation and protein secretion. *Inflamm Bowel Dis.* 20:1794.
 121. Zhang Y, Bertolino A, Fazio L, Blasi G, Rampino A, Romano R, et al. (2007): Polymorphisms in human dopamine D2 receptor gene affect gene expression, splicing,

- and neuronal activity during working memory. *Proc Natl Acad Sci U S A.* 104:20552-20557.
122. Niesler B, Flohr T, Nöthen MM, Fischer C, Rietschel M, Franzek E, et al. (2001): Association between the 5' UTR variant C178T of the serotonin receptor gene HTR3A and bipolar affective disorder. *Pharmacogenet Genomics.* 11:471-475.
 123. Pohjalainen T, Rinne J, Nägren K, Lehikoinen P, Anttila K, Syvälahti E, et al. (1998): The A1 allele of the human D-2 dopamine receptor gene predicts low D-2 receptor availability in healthy volunteers. *Mol Psychiatry.* 3:256-260.
 124. Chen H, Wilkins LM, Aziz N, Cannings C, Wyllie DH, Bingle C, et al. (2006): Single nucleotide polymorphisms in the human interleukin-1B gene affect transcription according to haplotype context. *Hum Mol Genet.* 15:519-529.
 125. Giedraitis V, He B, Huang W-X, Hillert J (2001): Cloning and mutation analysis of the human IL-18 promoter: a possible role of polymorphisms in expression regulation. *J Neuroimmunol.* 112:146-152.
 126. Haastруп E, Bukh JD, Bock C, Vinberg M, Thørner LW, Hansen T, et al. (2012): Promoter variants in IL18 are associated with onset of depression in patients previously exposed to stressful-life events. *J Affect Disord.* 136:134-138.
 127. Lachman HM, Nolan KA, Mohr P, Saito T, Volavka J (1998): Association between catechol O-methyltransferase genotype and violence in schizophrenia and schizoaffective disorder. *Am J Psychiatry.* 155:835-837.
 128. Chen Q-Y, Chen Q, Feng G-Y, Wan C-L, Lindpaintner K, Wang L-J, et al. (2006): Association between the brain-derived neurotrophic factor (BDNF) gene and schizophrenia in the Chinese population. *Neurosci Lett.* 397:285-290.
 129. Egan MF, Kojima M, Callicott JH, Goldberg TE, Kolachana BS, Bertolino A, et al. (2003): The BDNF val66met polymorphism affects activity-dependent secretion of BDNF and human memory and hippocampal function. *Cell.* 112:257-269.