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An exploratory study of combination buspirone and melatonin SR in Major Depressive Disorder (MDD): A possible role for neurogenesis in drug discovery[☆]

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ABSTRACT

We used *in vitro* neurogenesis-based human neural stem cell (hNSCs) assays and rodent *in vivo* behavioral assays to identify potential novel antidepressants. A combination of buspirone and melatonin displayed antidepressant activity in these assays whereas neither buspirone nor melatonin alone showed any antidepressant-like profile. After evaluating numerous combination ratios, we determined that low dose buspirone 15 mg combined with melatonin-SR 3 mg yielded optimal antidepressant efficacy in our pre-clinical platform. The low dose of buspirone suggested that antidepressant efficacy might be achieved with only minimal adverse event liability. Based on these data, we conducted an exploratory 6-week, multi-center, double-blind, randomized, placebo- and comparator-controlled study of the combination of buspirone and melatonin in subjects with acute Major Depressive Disorder (MDD). The combination treatment revealed a significant antidepressant response in subjects with MDD on several measures (Clinical Global Impression of Severity and Improvement, Inventory of Depressive Symptomatology) compared to either placebo or buspirone 15 mg monotherapy. These preliminary findings have clinical implications and suggest that a platform of pre-clinical neurogenesis matched with confirmatory behavioral assays may be useful as a drug discovery strategy.

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1. Introduction

Only 30% of patients with Major Depressive Disorder (MDD) achieve full remission of symptoms with currently available antidepressants (Blier and Ward, 2003; Rush et al., 2006; Trivedi et al., 2006; Rush, 2007; Nassir Ghaemi, 2008). These drugs are associated with adverse effects such as gastrointestinal symptoms, agitation, sleep disturbance, and sexual dysfunction that may affect treatment compliance. Consequently, there is still a clear need for antidepressant medications to treat MDD that possess enhanced efficacy, fewer adverse effects, and utilize different mechanisms of action than currently available medications (Bosker et al., 2004; Nassir Ghaemi, 2008; Rush et al., 2008). Historically, drug

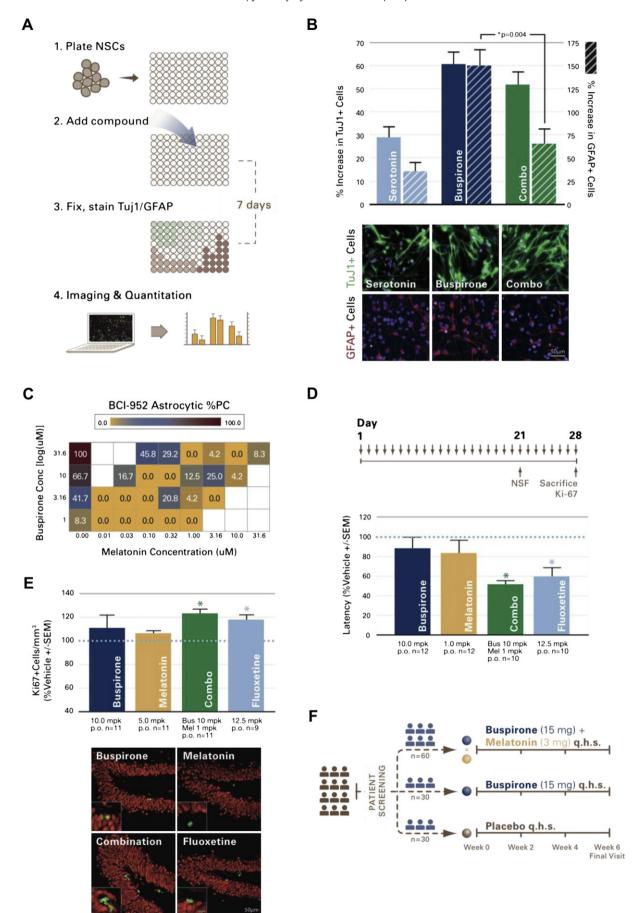
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development relied upon extrapolations from the monoamine hypothesis or serendipity (Bosker et al., 2004; Kasper and Hamon, 2009), but some recent strategies have employed more rational pre-clinical foundations to investigate novel mechanisms of action.

One type of strategy to facilitate drug development for mood disorders may be a pre-clinical platform of neurogenesis-based assays (Aimone et al., 2010). It has been demonstrated that neural stem cells in the adult human brain undergo neurogenesis, and that the resulting new cells are integrated into neural circuits that can become fully functional neurons (Eriksson et al., 1998). Several studies have specifically linked hippocampal function and neurogenesis to mood disorders (Jacobs et al., 2000; Drevets, 2001; van Praag et al., 2002; Gold and Chrousos, 2002; Warner-Schmidt and Duman, 2006; Sahay and Hen, 2007; Boldrini et al., 2009; David et al., 2009; Lucassen et al., 2010; Dranovsky et al., 2011). It has been reported that reduced hippocampal volume in untreated depression may be reversed by treatment with antidepressant medications, and that blocking neurogenesis makes all FDA-approved antidepressants ineffective (Malberg et al., 2000; Sheline et al., 2003; Santarelli et al., 2003; Dranovsky and Hen, 2006; Sahay

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and Hen, 2007; Wang et al., 2008; Boldrini et al., 2009). These findings suggest that hippocampal neurogenesis may be implicated in the expression of depressive symptoms. It is important to note that the role of neurogenesis in the etiology of and recovery from depression is still not clear (Henn and Vollmayr, 2004; Gass and Henn, 2009; Hanson et al., 2011; Aimone et al., 2011). Besides mood, many other factors including stress, enriched environments, and exercise can influence hippocampal neurogenesis (Boldrini et al., 2009; Lucassen et al., 2010; Snyder et al., 2011; Hanson et al., 2011. Therefore, it is premature to draw any conclusions about the specific role despite strong support for some role for neurogenesis in depression. Nevertheless, the evidence to date does point to a different, intriguing approach for rational drug development for mood disorders based upon the pharmacological induction of neurogenesis (Sahay and Hen, 2007; Aimone et al., 2010; Duman and Voleti, 2012).

BrainCells Inc. (BCI) developed an in vitro and in vivo neurogenesis platform to facilitate the identification of novel antidepressant treatment strategies, including drug combinations. Using this rationale for drug discovery, BCI studied a broad matrix of nonproprietary drugs to seek optimal drug combinations. Once a drug combination was identified, pharmacokinetic and dose optimization studies were conducted to identify the optimal dose range to maximize clinical efficacy and minimize the adverse event liability. Using this strategy, the combination of buspirone and melatonin was identified. This drug combination demonstrated both enhanced neurogenesis in the pre-clinical neurogenesis-based in vitro assays and antidepressant responses in the in vivo behavioral assays that were equivalent to a Serotonin Selective Reuptake Inhibitor (fluoxetine). Further, buspirone was effective in these assays at doses below those generally used in clinical practice for Generalized Anxiety Disorder (GAD) and well below doses reported for antidepressant efficacy in the literature (Schweizer et al., 1986; Robinson et al., 1990; Blier and Ward, 2003; Rickels et al., 2003). The lower dose of buspirone made the buspirone-melatonin combination of particular interest because of its potential clinical utility.

Buspirone is a partial 5-hydroxytryptamine 1_a (5-HT1_a) receptor agonist as well as a presynaptic dopamine antagonist (D₂) and partial alpha_{1,2} receptor antagonist (Deakin, 1993; Blier et al., 1997; Stahl et al., 1998; Adeagbo et al., 2000). The potential of 5-HT1_a receptor agonism has been studied extensively in MDD. A lack of

normal responsiveness of the post-synaptic 5-HT1a receptor is implicated in the reduced level of serotonin available in the synapse as well as stimulation of pre-synaptic somatodendritic 5-HT1a receptors which inhibits serotonin synthesis and release (Deakin, 1993; Stahl et al., 1998; Blier and Ward, 2003). 5-HT1a agonists like buspirone, gepirone, and ipsapirone have been evaluated in several double-blind, placebo-controlled trials of MDD but have never received FDA approval (Rickels et al., 1990; Robinson et al., 1990; Rickels et al., 1991; Sramek et al., 1996; Wilcox et al., 1996; Heiser and Wilcox, 1998; Lapierre et al., 1998; Stahl et al., 1998; Alpert et al., 2004; Amsterdam et al., 2004; Keller et al., 2005; Bielski et al., 2008). A few studies suggested that buspirone doses above 40 mg per day were efficacious for acute MDD (Schweizer et al., 1986; Robinson et al., 1990). In fact, Blier and Ward (2003) contended that higher doses of azapirones were required in order to demonstrate efficacy in depression. However, a higher dose of buspirone has not been clinically practical because it can cause dizziness, drowsiness, headache, fatigue, nervousness, insomnia, lightheadedness, cognitive impairment, as well as nausea, gastric distress, palpitations, nightmares, tremor, numbness (paresthesia), and blurred vision (Rickels et al., 2003). Consequently, further development of buspirone as an antidepressant has been restricted by the adverse évent liability at higher doses.

Combination or augmentation treatment strategies have been endorsed by many clinicians and were employed as a fundamental part of the NIMH-sponsored sequenced treatment alternatives to relieve depression study, STAR*D (Thase et al., 1998; Fava and Rush, 2006; Rush, 2007). Buspirone has been used effectively as adjunctive treatment with SSRI's in patients with MDD (Landen et al., 1998; Appelberg et al., 2001; Trivedi et al., 2006; Rush, 2007). Of course, the addition of buspirone to SSRI's still conveys the same adverse event liabilities as SSRI monotherapy. Therefore, combining 5-HT1_a agonists with an *alternative* medication to the SSRI's might be of clinical value. Ideally, a combination/augmentation strategy would allow lower doses of buspirone to be used in order to minimize the adverse event potential of the azapirones as well.

The FDA approved busiprone for the treatment of Generalized Anxiety Disorder (GAD) in 1986. The typical recommended dose of buspirone ranges between 20 and 60 mg/day in two divided doses, with 30 mg/day generally considered as the typical therapeutic dose for GAD (Rickels et al., 2003). As noted above, the pre-clinical

Fig. 1. Pre-clinical drug discovery platform of human in vitro and rodent in vivo assays with buspirone and melatonin to explore neuronal differentiation and antidepressant behavioral effects. A) Depiction of hNPC experimental flow: hNPC neurospheres were dissociated, plated in 96-well plates and allowed to differentiate for seven days in the presence of test compounds. Cells were fixed and stained for Tu[1, GFAP and Hoechst, imaged, quantitated and analyzed for increases in neuronal or glial differentiation. B) Effect of serotonin, buspirone, and the combination of buspirone and melatonin on the differentiation of hNPC's in vitro: Cells staining positive for TuJ1 or GFAP were manually counted on a field-byfield basis as described in the methods. The percent increase in TuJ1 positive cells is indicated on left sided Y-axis (solid bar) and on the right side for GFAP positive cells (hatched bar). Effects of serotonin are indicated in light blue, buspirone in dark blue, and the combination in green. The lower panels are representative images as indicated (Combo = buspirone and melatonin). The neuronal marker TuJ1 is shown in green (upper images), the glial marker GFAP is shown in red (lower images) and the nuclear stain Hoechst is shown in blue in both upper and lower panels. The scale bar indicates 50 µM. C) Heatmap representing the in vitro effects of varying concentration and ratios of melatonin on blocking buspirone mediated induction of GFAP positive cell formation: Scale is indicated by color ranging from red (high GFAP density) to yellow (low GFAP density). Y-axis represents increasing concentrations of buspirone and the x-axis represents increasing concentrations of melatonin. D) Activity of the combination of buspirone and melatonin in NSF: Experimental design for assessing the effect of compounds on NSF and neurogenesis is shown in the upper figure. Arrows at the top indicate daily oral dosing of drugs or vehicle. Arrows at the bottom indicate experimental day of novelty suppressed feeding (NSF) assay or animal sacrifice, perfusion/fixation and subsequent immunohistochemistry. Shown in the lower panel is the NSF assay data as a % of appropriate vehicle control (Latency, % Vehicle Y-axis). Treatment and dose in milligram per kilogram body weight (mpk) is indicated on the X-axis with fluoxetine (light blue), buspirone (dark blue), melatonin (yellow) or the combination of buspirone and melatonin (Combo-green). n indicates number of animals in each group. *indicates p < 0.01 for combination treatment and p < 0.05 for fluoxetine alone based on an unpaired two-tailed Student's t-test comparing drug treatment versus appropriate vehicle control. E) Change in the number of NSC as assessed by Ki67 staining is shown as the total number of Ki-67-positive cells per cubic mm (Ki-67+ Cells/mm³) within the dentate gyrus expressed as a % of appropriate vehicle control after 28 days of treatment. Dosing of rats was as described for Fig. 1D. Effects on Ki-67 positive cells are shown for fluoxetine (light blue), buspirone (dark blue), melatonin (yellow) and the combination of buspirone and melatonin (Combo-green). n indicates number of animals in each group. * indicates p < 0.01 and *p < 0.05 value based on an unpaired two-tailed Student's t-test comparing drug treatment versus appropriate vehicle control. The lower panels are representative images taken at 40× magnification using a confocal microscope. Insets are shown at 2× magnification. The mature neuronal marker NeuN is shown in red (indicating the granule cell layer of the dentate gyrus), and cells labeled with Ki-67 are shown in green. The scale bar indicates 50 μM. F) Diagram of the clinical study: Shown is a schematic of the study design. After successful completion of screening procedures, approximately 120 eligible patients were randomized at week 0 to one of three study treatment groups at a ratio of 2:1:1. The top line labeled buspirone (15 mg) + melatonin (3 mg) represents the combination treatment group; the buspirone (15 mg) alone and the placebo groups are shown on the lines below. Study treatment was administered once a day at bedtime (q.h.s). All patients were evaluated every 2 weeks and the duration of the trail was 6 weeks.

neurogenesis-based data suggesting that buspirone doses *below* 20 mg combined with melatonin might yield an antidepressant effect has broad clinical implications. Consequently, we conducted a clinical trial to determine if this low dose buspirone—melatonin combination might yield antidepressant efficacy and minimize the dose-related side effects associated with higher doses of buspirone.

2. Methods

2.1. Pre-clinical drug discovery and dose optimization

We used neurogenesis-based in vitro screening assays using human neural progenitor cells (hNPCs) and specific rodent behavioral in vivo assays to create a unique pre-clinical platform for drug discovery (Appendix 1). Human NPCs isolated from intact brain grown as primary cells in culture retain their ability to produce additional progenitor cells for a restricted time (Fig. 1A). These hNPCs cells can be stimulated to differentiate into mature cells that phenotypically resemble neurons and express an early neuronal marker β-Tubulin III (Tuj1) or into cells that express markers for non-neuronal lineages, like astrocytes (Glial Fibrillary Acidic Protein-GFAP) in the absence of human serum. Exposure of hNPCs to serotonin increases the number of cells that differentiate into neurons consistent with a role for serotonin in promoting neurogenesis (Malberg et al., 2000; Sahay and Hen, 2007; Wang et al., 2008). Drugs like fluoxetine are very neurogenic in this assay platform. On the other hand, we found that buspirone caused hNPC's to differentiate into both neuronal and non-neuronal lineages (Fig. 1B). We postulated that the increased formation of nonneuronal cells might adversely affect the putative antidepressant efficacy of buspirone. Consequently, we sought a non-toxic drug that in combination with buspirone would suppress the buspironeinduced GFAP response while sustaining the buspirone-induced neurogenesis response. BCI examined over 60 different drugs in combination with buspirone to identify the best combination. We found that the addition of melatonin markedly reduced the nonneuronal differentiation produced by buspirone alone (Fig. 1B), although melatonin alone had no effect on hNPCs at all.

Subsequently, we evaluated a range of buspirone—melatonin combination ratios *in vitro* (Fig. 1C). Within the selected dosage ranges, a combination ratio of 1:10 (melatonin to buspirone) appeared to sufficiently suppress GFAP responses while sustaining the neurogenesis response.

Using the combination ratio derived from the in vitro assays as a guide, we applied these findings to determine dosing for the in vivo novelty suppressed feeding (NSF) assay. We aimed for the lowest possible dose range in animals and focused on achieving a mg/kg animal dosage (mpk) that could be scaled to clinically relevant human doses. The NSF assay is a conflict assay that assesses the balance between an animal's desire to approach a palatable reward and a fear of entering an open space (Santarelli et al., 2003; Dranovsky and Hen, 2006). When administered chronically (but not acutely), antidepressants are typically active in the NSF assay, whereas inhibition of hippocampal neurogenesis blocks the activity of antidepressants (Santarelli et al., 2003). Fisher 344 rats were chronically dosed with the optimized buspirone melatonin combination ratio in the NSF assay and studied for changes in behavior and hippocampal neurogenesis (Fig. 1D and E). Neither buspirone nor melatonin alone showed efficacy in either NSF or histological assessment of neurogenesis (examining the number of NSCs as assessed by Ki67 staining). However, the selected drug combination showed activity comparable to the marketed antidepressant fluoxetine on both the NSF assay (Fig. 1D) and on histological analysis (Fig. 1E).

It is well known that dose selection based upon either *in vitro* assays or *in vivo* animal studies have serious limits and can only approximate human dosing possibilities. Ultimately, using the preclinical neurogenesis data and available species comparisons of AUC (Area under the curve) and plasma $C_{\rm max}$ values (maximum plasma concentration), we selected 3 mg of melatonin (sustained release) and 15 mg of buspirone as best dose estimates. The buspirone dose is *below* the typical recommended therapeutic range for anxiety and well below the previously reported 40–60 mg effective doses for MDD (Robinson et al., 1990; Rickels et al., 2003). Clearly, the lower buspirone dose suggests that an intriguing clinical benefit might be gained for patients with MDD. The typical clinical dose of melatonin ranges from approximately 1 to 10 mg/day.

2.2. Clinical trial methods

This nine-site study was conducted in the United States as an Investigator-Initiated Phase 2 Clinical Trial coordinated by Massachusetts General Hospital (MGH) and sponsored by BrainsCells Inc. (BCI, San Diego, California) which supplied study drug and matching placebo capsules. The protocol and informed consent forms were approved by a central Ethics Committee or by the Ethics Committee commonly used by the participating Investigative site.

2.2.1. Study participants

Subjects between 18 and 65 years of age who met DSM-IV-TR criteria for Major Depressive Disorder (MDD), as determined by the Mini-International Neuropsychiatric Interview (M.I.N.I.) and psychiatric evaluation were eligible for this study (Sheehan et al., 1998). All subjects gave written documentation of informed consent before any study related procedures. Eligible subjects required a total score of 14 or higher on the 16-item Quick Inventory of Depressive Symptoms-Self-Rated scale (QIDS-SR₁₆)) at both the screening and baseline visits (Rush et al., 2003).

Female patients of childbearing potential were on a reliable, medically acceptable form of contraception for at least 30 days prior to screening and throughout the study. Subjects meeting criteria for other Axis-I disorders as their primary diagnosis, had a history of eating disorders, obsessive-compulsive disorder, psychotic disorder, bipolar disorder and/or mental retardation and those with alcohol or substance abuse or dependency were excluded. Use of antidepressant, antipsychotic, or anxiolytic medication or drugs with known psychotropic properties were prohibited for 1 week (4 weeks for fluoxetine) prior to screening and throughout the study. Subjects who used substances that are known inhibitors or inducers of CYP3A4 were excluded.

2.2.2. Study design and randomization procedure

This was a double-blind, randomized, placebo-controlled study using a 2:1:1 allocation between 3 treatment groups using a computer-generated centralized randomization schedule. Subjects received either: the study drug combination of buspirone immediate release (IR) 15 mg with melatonin slow release (SR) 3 mg; buspirone IR 15 mg as monotherapy; or matching placebo capsules for 6 weeks following randomization (Fig. 1F). Buspirone was obtained from Bristol Myers Squibb, melatonin SR was obtained from Mellen Medical Products, Inc. and the clinical trial material was prepared by Fisher Clinical Services.

Buspirone IR 15 mg tablets and melatonin SR 3 mg tablets were over-encapsulated for blinding purposes. Subjects took two capsules of study drug once a day at bedtimewith or without food. Subjects, investigators, study staff and BCI personnel involved in the trial were blinded to study group assignment.

2.2.3. Study procedures

After subjects signed the informed consent, a M.I.N.I. diagnostic and psychiatric screening evaluation was done in addition to measurement of vital signs and routine blood and urine tests. The QIDS-SR₁₆ and Clinical Global Impression of Severity (CGI-S) were administered at screening, baseline and after 2, 4, and 6 weeks of treatment and the CGI-Improvement (CGI-I) was done after 2, 4, and 6 weeks (Rush et al., 2003; Guy, 1976; Targum et al., 2008). Additional efficacy assessments included the 30-item Inventory of Depressive Symptomatology-Clinical version (IDS-C₃₀) and the Hamilton Rating Scale for Anxiety (HAM-A) that were done at baseline and after 6 weeks (Hamilton, 1959; Rush et al., 1996). Adverse events were recorded at each study visit.

2.2.4. Outcome measures

The primary endpoint in this exploratory study was the mean change from baseline of the CGI-I after 6 weeks of study drug treatment or the last available post-randomization CGI-I score available. Additional endpoints included change from baseline in the CGI-S, QIDS-SR₁₆, IDS-C₃₀ and HAM-A. In addition, responder analysis was carried out for the CGI-I and responder and remission analyses for the IDS-C₃₀ on the modified intent to treat (MITT) population. The CGI-I response was defined all subjects obtaining a score of 1 or 2. The QIDS-SR₁₆ and IDS-C₃₀, response was defined as subjects obtaining scores less than or equal to half of their baseline score. Clinical remission was defined as an endpoint score of \leq 5 for the QIDS-SR₁₆ and \leq 11 for the IDS-C₃₀.

Safety assessments were based upon evaluation of vital signs and adverse events reported during the study.

2.2.5. Statistical analysis

Statistical analyses included Mixed-effects Model Repeated Measures (MMRM) for instruments that were assessed at multiple time points (CGI-S, CGI-I, QIDS-SR₁₆) in the MITT population using PROC mixed within SAS (Lieberman et al., 2005). In these analyses, the available data from each patient was used, without any imputation for missing values. Alternatively, for measures performed only

at baseline and week 6 (IDS- C_{30} and HAM-A), the efficacy analyses used an analysis of covariance (ANCOVA) model, with change from baseline as the dependent variable, the baseline value as a covariate, and treatment group as the factor with three. Point estimates represent the model-based estimate of the mean for this sampling of the population. A responder analysis was calculated for the CGI-I and responder and remission analyses for the IDS- C_{30} and QIDS-SR $_{16}$ on the MITT population. Fisher's Exact Test was used to determine the significance level of differences in response and remission rates across treatments. Paired t-tests were used to determine significance of changes in vital signs during the treatment phase of the study.

By design, the planned statistical analyses for this small study included a secondary pooling of the buspirone and placebo treatment groups on the expectation that these groups would not differ on the mean CGI-I at endpoint by more than 0.04 points.

3. Results

A total of 142 subjects consented, enrolled, and were randomized in the study. Eight subjects were randomized but never received study drug. The MITT and Safety populations were identical and consisted of 134 randomized patients (33 patients in the placebo group, 34 patients in the buspirone group, and 67 patients in the combination treatment group). 112 patients who received study drugs (83.6%) completed the study.

Demographic and baseline characteristics were similar among treatment groups (Table 1). The study population was predominantly female (64.9%). Patients had a mean age of 42.4 years, mean height of 66.3 inches, and a mean weight of 203.9 pounds. There were no differences in demographics across treatment groups.

There were no statistically significant differences between the baseline efficacy measures between treatment groups (Table 1).

3.1. Efficacy outcomes

The mean efficacy scores at baseline, point change estimates for CGI-I. and the mean changes from baseline for the other efficacy

 Table 1

 Baseline characteristics of modified intent to treat population.

Variable	Placebo (P) $(n = 33)$	Buspirone (B) $(n = 34)$	Combination $(B + M)^a$ $(n = 67)$	Overall ($n=134$)
Age (years)				
N	33	34	67	134
Mean (SD)	42.7 (11.62)	40.8 (12.55)	43.1 (12.06)	42.4 (12.03)
Median	43.9	40.7	44.7	43.9
Min, max	23.4, 65.0	19.8, 62.7	19.3, 64.8	19.3, 65.0
Gender (n [%])				
Female	24 (72.7)	20 (58.8)	43 (64.2)	87 (64.9)
Male	9 (27.3)	14 (41.2)	24 (35.8)	47 (35.1)
Ethnicity (n [%])				
Latino	5 (15.2)	5 (14.7)	8 (11.9)	18 (13.4)
Not Latino	28 (84.8)	29 (85.3)	59 (88.1)	116 (86.6)
Race				
Asian	1 (3.0)	2 (5.9)	1 (1.5)	4 (3.0)
American Indian	1 (3.0)	0	0	1 (0.7)
Black	14 (42.4)	16 (47.1)	27 (40.3)	57 (42.5)
White	16 (48.5)	15 (44.1)	34 (50.7)	65 (48.5)
Other	1 (3.0)	1 (2.9)	5 (7.5)	7 (5.2)
QIDS-SR ₁₆				
Mean (SD)	17.24 (3.307)	16.88 (2.306)	16.98 (3.079)	17.02 (2.945)
Median	16	16	16	16
Min, Max	14, 24	14, 23	11 ^b , 25	11 ^b , 25

a Combination (B + M) indicates the group treated with both buspirone and melatonin.

b One patient who had a QIDS-SR₁₆ score below the entry criteria was mistakenly enrolled and randomized to the combination treatment group.

Table 2Baseline and endpoint analysis of efficacy instruments: MITT population.

Outcome measure	Combination (B + M) ^a		Placebo		B + M ^a vs. placebo	Buspirone		B + M ^a vs. buspirone	B + M ^a vs. pooled placebo and buspirone
	Mean baseline	Adjusted change	Mean baseline	Adjusted change	p Value	Mean baseline	Adjusted change	p Value	p Value
CGI-I ^b	n/a (n = 67)	2.37 (n = 67)	n/a (n = 33)	2.86 (n = 33)	0.046	n/a (n = 34)	2.82 (n = 34)	0.068	0.021
CGI-S ^c	4.51 (n = 67)	1.43 (n = 67)	4.45 (n = 33)	0.93 (n = 33)	0.04	4.53 (n = 34)	0.88 (n = 34)	0.02	0.009
QIDS-SR ₁₆ ^c	16.94 (n = 67)	8.57 (n = 67)	17.24 (n = 33)	7.31 (n = 33)	0.24	16.88 (n = 34)	6.65 (n = 34)	0.08	0.07
IDS-C ₃₀ d	40.66 (n = 67)	19.24 (n = 54)	40.27 (n = 33)	14.42 (n = 30)	0.034	41.76 (n = 34)	16.35 (n = 28)	0.155	0.031
HAM-A ^d	19.55 (n=67)	9.04 (n = 54)	20.42 (n = 33)	6.51 (n = 30)	0.041	20.00 (n = 34)	6.69 (n = 28)	0.136	0.032

- (n) indicates the number of subjects for each measurement.
 - $^{
 m a}$ Combination (B + M) indicates the group that was treated with both buspirone and melatonin.
- b CGI-I value for baseline is not applicable (n/a) as there is no baseline score for this assessment. Adjusted change indicates point estimates calculated from the MMRM on the MITT population and p-values are based on the MMRM on the MITT population.
- ^c CGI-S and QIDS-SR₁₆ mean baseline scores are calculated as arithmetic means from all patients in the MITT population. Adjusted change indicates point estimates calculated from the MMRM on the MITT population and *p*-values are based on the MMRM on the MITT population.
- d IDS-C₃₀ and Ham-A mean baseline scores are calculated as arithmetic means on all subjects in the MITT population. Adjusted change indicates the ANCOVA derived change for those patients who completed both the baseline and endpoint evaluations (completers) as the instruments were assessed at baseline and endpoint only.

measures at week 6 by treatment group are presented in Table 2. The CGI-I improved significantly in the combination group compared to placebo (MMRM; p=0.047). Similarly, the CGI-S revealed statistically significant improvement in the combination treatment compared to either buspirone monotherapy (MMRM; p=0.02) or placebo (MMRM; p=0.04). Changes from baseline to week 6 in the QIDS-SR₁₆, IDS-C₃₀, and HAM-A also revealed clinical benefit for the combination treatment relative to the buspirone monotherapy or placebo groups. The combination treatment was significantly better than placebo on the IDS-C₃₀ (ANCOVA; p=0.034), but not on the QIDS-SR₁₆.

Comparison between the buspirone 15 mg monotherapy and placebo groups revealed no clinical differences on any measure.

The mean endpoint CGI-I scores were 2.33 \pm 0.97 (SD) for the combination treatment group, 2.79 \pm 1.24 for the buspirone 15 mg group, and 2.83 \pm 1.09 for the placebo group. The statistical analysis plan included a secondary pooling of the buspirone and placebo treatment groups if their respective mean CGI-I scores at endpoint were no greater than 0.04 points apart. In fact, the mean difference for the CGI-I endpoints for the buspirone and placebos groups was only 0.04. As revealed in Table 2, the pooling of these two groups enhanced the statistical significance in favor of the combination treatment.

The CGI-I response rate was 58.2% in the combination group, 38.4% in the buspirone monotherapy group, and 36.4% in the placebo group. The advantage for the combination treatment approached statistical significance for both the buspirone (p=0.00.06) and placebo groups (p=0.055). When the buspirone monotherapy and placebo groups are pooled versus the combination treatment, the difference does reach statistical significance (p=0.024).

As shown in Table 3, response and remission rates were also calculated for the IDS- C_{30} . and QIDS- SR_{16} . These remission rates reveal that the combination treatment was significantly better than buspirone 15 mg monotherapy on both the IDS- C_{30} (p=0.045) and the QIDS- SR_{16} (p=0.0078).

There was a significant improvement from baseline to week 6 on the mean Ham-A scores in the combination treatment group compared to placebo (p=0.04) and a non-significant trend for buspirone alone (p=0.14). Alternatively, buspirone 15 mg monotherapy revealed no clinical difference on the HAM-A compared to placebo (p=0.62) reflecting a no anxiolytic benefit at that low dose.

3.2. Safety and tolerability

30 subjects did not complete the study of who eight never received study drug. Among the non-completers, only eight

subjects withdrew from the study due to adverse events, 6% in each of the treatment groups. There were no significant group differences noted amongst the 22 non-completers who received study drug.

The combination treatment was well tolerated; the frequency of treatment-emergent adverse events (TEAEs) was similar between the treatment groups (Table 4). No serious TEAEs were reported.

Overall, 52.2% of the subjects in this study experienced some treatment emergent adverse events (TEAEs). 51.5.5% of subjects in the placebo group, 61.8% in the buspirone group, and 47.8% in the combination treatment group had TEAE's. Of the subjects experiencing TEAEs, 35.8% of these events were considered study drug related (33.3% of subjects in the placebo group, 44.1% in the buspirone group, and 32.8% in the combination treatment group). 15.7% of the TEAEs were assessed as mild in intensity, 16.4% as moderate in intensity, and 3.7% were assessed as severe in intensity.

4. Discussion

In this small exploratory study of subjects with acute MDD, low dose buspirone (15 mg) combined with melatonin-SR 3 mg yielded a significant antidepressant effect compared to placebo or buspirone monotherapy during 6-weeks of double-blind treatment. 58.2% of MDD subjects treated with the combination were CGI-I responders in contrast to only 36.4% of placebo subjects and 38.2% of subjects receiving only buspirone 15 mg. Similarly, significantly positive findings were noted for the CGI-S, IDS-C₃₀, and HAM-A. These efficacy results compare favorably with reported results from other acute depression studies with approved antidepressant compounds and the STAR*D study (Trivedi et al., 2006; Rush et al., 2008). In retrospect, it would have been informative to include the Hamilton rating scale for depression (Ham-D) in the psychometrics as well.

The combination of buspirone and melatonin was well tolerated and safe. An analysis of the sleep items from the QIDS-SR₁₆ revealed no significant difference in response between subjects who received the combination or placebo. This finding suggests that a possible melatonin effect on sleep was not the basis for the clinical improvement in the depressed subjects.

This was a small double-blind, placebo-controlled proof-of-concept study. Buspirone was included as a third, separate treatment group to confirm the expected lack of antidepressant effect of this low dose. In fact, buspirone 15 mg monotherapy was not differentiated from placebo for depressive or anxiety symptoms in this study. Melatonin was not added as a fourth treatment group

Table 3Responder and remission Analyses: MITT population.

Responder variable	n	Responders (%)	Buspirone vs. placebo <i>p</i> -value	Combination $(B + M)^a$ vs. placebo p -value	$(B + M)^a$ vs. buspirone p -value	(B + M) ^a vs. pooled placebo & buspirone <i>p</i> -value
CGI-I at week 6						
Placebo	33	12 (36.36%)	1.0000	0.0554	0.0627	0.0242
Buspirone	34	13 (38.24%)				
Combination $(B + M)^a$	67	39 (58.21%)				
QIDS-SR ₁₆ at week 6						
Placebo	33	14 (42.42%)	0.8056	0.5245	0.2927	0.2979
Buspirone	34	13 (38.24%)				
Combination $(B + M)^a$	67	34 (50.75%)				
IDS-C ₃₀ at week 6						
Placebo	32	10 (31.25%)	0.6029	0.1275	0.5053	0.1465
Buspirone	31	12 (38.71%)				
Combination $(B + M)^a$	60	29 (48.33%)				
QIDS-SR ₁₆ at week 6 remis	ssion					
Placebo	33	8 (24.24%)	0.1092	0.4879	0.0078	0.0440
Buspirone	34	3 (8.82%)				
Combination $(B + M)^a$	67	22 (32.84%)				
IDS-C _{.30} at week 6 remissi	on					
Placebo	32	6 (18.75%)	0.2565	0.6062	0.0453	0.1058
Buspirone	31	2 (6.45%)				
Combination $(B + M)^a$	60	15 (25.00%)				

Fisher's Exact Test was used to compare responses between groups for the responder and remission analyses. CGI-I responders had scores of 1 or 2. QIDS-SR₁₆, and IDS-C₃₀, responders had \geq 50% improvement from baseline. QIDS-SR₁₆ remission was defined as scores \leq 5 and IDS-C₃₀ remission as scores \leq 11.

because it is not known to have antidepressant properties. However, the absence of a melatonin comparator is still a limitation in the interpretation of the results.

The potential that 5-HT1_a agonists might have antidepressant properties is not a new finding. Previous studies have suggested that doses of buspirone above 40 mg had antidepressant effects in MDD (Robinson et al., 1990; Rickels et al., 1991), and both gepirone and ipsapirone have been studied in MDD (Sramek et al., 1996; Wilcox et al., 1996; Blier et al., 1997; Heiser and Wilcox, 1998; Lapierre et al., 1998; Stahl et al., 1998; Alpert et al., 2004; Amsterdam et al., 2004; Keller et al., 2005; Bielski et al., 2008). Recently, vilazodone, an SSRI and partial 5-HT1_a agonist was

approved by the FDA (Rickels et al., 2009; Laughren et al., 2011). Regarding melatonin, agomelatine reveals melatonin receptor 1 and 2 partial agonism as well as serotonin 2C receptor antagonism. There have been several preclinical and clinical studies demonstrating the efficacy of agomelatine as a neurogenic agent and as a treatment for MDD (Eser et al., 2007; Kasper and Hamon, 2009; Bourin and Prica, 2009; Soumier et al., 2009). Of course, there are key differences in the receptor pharmacology between agomelatine and the combination of buspirone and melatonin. Buspirone is an agonist on the serotonin 1A receptor, does not affect serotonin 2C, and melatonin has agonistic properties at all known melatonin receptors including MT3 in addition to MT1 and MT2.

Table 4Summary of TEAE's, including TEAEs reported in >5% of patients and higher than Placebo: Safety population.

TEAEs [n (%)]	Placebo ($n=33$)	Buspirone 15 mg ($n = 34$)	Combination $(B + M)^a$ $(n = 67)$	Overall (<i>n</i> = 134)
Patients with any TEAE	17 (51.5%)	21 (61.8%)	32 (47.8%)	70 (52.2%)
Patients with any TEAE related to drug	11 (33.3%)	15 (44.1%)	22 (32.8%)	48 (35.8%)
Patients with serious TEAEs	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)
Patients discontinued study due to TEAEs	2 (6.1%)	2 (5.9%)	4 (6.0%)	8 (6.0%)
Commonly occurring TEAEs				
Gastrointestinal disorders	7 (21.2%)	8 (23.5%)	14 (20.9%)	29 (21.6%)
Diarrhea ^b	2 (6.1%)	1 (2.9%)	6 (9.0%)	9 (6.7%)
Constipation ^c	0 (0.0%)	2 (5.9%)	1 (1.5%)	3 (2.2%)
Nervous system disorders	6 (18.2%)	7 (20.6%)	11 (16.4%)	24 (17.9%)
Dizziness ^b	0 (0.0%)	1 (2.9%)	4 (6.0%)	5 (3.7%)
Headache ^{b,c}	2 (6.1%)	3(8.8%)	5(7.5%)	10(7.5%)
TEAEs of special interest				
Dry mouth	1 (3.0%)	1 (2.9%)	2 (3.0%)	4 (3.0%)
Nausea	2 (6.1%)	1 (2.9%)	4 (6.0%)	7 (5.2%)
Insomnia	0 (0.0%)	1 (2.9%)	1 (1.5%)	2 (1.5%)
Middle insomnia	1 (3.0%)	0 (0.0%)	0 (0.0%)	1 (0.7%)
Terminal insomnia	0 (0.0%)	0 (0.0%)	1 (1.5%)	1 (0.7%)

Note: Related = definitely, probably, or possibly related to study drug and the n indicates the number of subjects in each group.

 $^{^{}m a}$ Combination (B + M) indicates the group that was treated with both buspirone and melatonin.

 $^{^{\}rm a}$ Combination (B + M) indicates the group that was treated with both buspirone and melatonin.

 $^{^{}m b}$ TEAEs occurring in >5% of patients in the combination (B + M) treatment group and higher than placebo.

^c TEAEs occurring in >5% of patients in the buspirone group and higher than placebo.

The identification and dose optimization of the combination of buspirone and melatonin was facilitated by a preclinical neurogenesis-based drug discovery platform. The *in vitro* assays guided the selection and ratio of the combination for *in vivo* NSF assays. In turn, these pre-clinical studies informed the selection of clinically relevant doses for clinical trials.

The finding that adult humans retain the ability to generate new neurons in the dentate gyrus of the hippocampus throughout life offers the potential for therapeutic intervention (Eriksson et al., 1998). However, the specific role of hippocampal neurogenesis in the etiology or resolution of depression is not clear. The preclinical finding that all classes of marketed antidepressants promote an increase in neurogenesis specific to the dentate gyrus is consistent with the concept that enhancing neurogenesis in humans may be clinically meaningful (Dranovsky and Hen, 2006; Warner-Schmidt and Duman, 2006; Aimone et al., 2010). Further, suppressing neurogenesis blocks some of the effects of antidepressants in the *in vivo* NSF assay (Santarelli et al., 2003; Li et al., 2008). Hence, there appears to be a strong link between antidepressant efficacy and neurogenesis (Surget et al., 2008; David et al., 2009; Dranovsky et al., 2011). However, some recent studies imply that neurogenesis may not be involved at all (Henn and Vollmayr, 2004; Gass and Henn, 2009; Hanson et al., 2011; Aimone et al., 2011). It is known that adult neurogenesis also decreases with stress in rodents and is enhanced by environmental enrichment or exercise, as well as the introduction of antidepressant medications (Boldrini et al., 2009; Lucassen et al., 2010; Dranovsky et al., 2011; Hanson et al., 2011). Further, animal studies do not always correlate neurogenesis levels with behavioral measurements of affective status (Holick et al., 2008; Miller et al., 2008). Boldrini et al. (2009) has suggested a broader view in that the observed impaired hippocampal plasticity associated with the pathogenesis of MDD is not merely due to impaired neurogenesis but to impaired cell connectivity and functional integration of brain circuitry regulating emotional responses as well.

Clearly, these preliminary clinical findings do *not* prove that neurogenesis is implicated in the antidepressant efficacy that was achieved in the trial. However, given the above limitations, it is still conceivable that the antidepressant effect of the combination treatment was achieved, in part, through sustained buspirone-induced neuronal proliferation and reversal by melatonin of the buspirone-induced stimulation of non-neuronal lineages (as measured by GFAP).

The precise nature of antidepressant-induced neurogenesis remains difficult to define, and most studies have relied on measures of proliferation assessed by incorporation of 5'-Bromo-2deoxyuridine (BrdU) or detection of Ki-67 in cells localized to the dentate gyrus and by confocal analysis for neuronal differentiation. These studies show that the majority (80–90%) of neural stem cells (NSCs) become neurons (Brown et al., 2003; Snyder et al., 2009). Formation of cell types other than neurons in vivo is difficult to assess, but newly developed in vitro techniques using human NPCs allowed us to study the effects of compounds on a variety of cell fates (Svendsen et al., 1998; Dranovsky et al., 2011). Exposure of hNPCs to serotonin increases the number of cells that differentiate into neurons, consistent with a role for serotonin in promoting neurogenesis (Wang et al., 2008). In contrast, buspirone increased not only the number of neuronal-positive cells but also the number of non-neuronal (GFAP positive) cells. Melatonin repressed buspirone-induced, GFAP-positive cell formation without eliminating the ability of buspirone to promote neuronal differentiation. Melatonin had no effect in the assays alone suggesting that melatonin works synergistically specifically with buspirone. Our data does not rule out that there are subtle effects of melatonin on neurogenesis in some settings and in certain experimental paradigms (Ramirez-Rodriguez et al., 2009). The combination of buspirone and melatonin had a robust effect in the NSF assay, whereas neither agent alone was effective. The combination also resulted in an increase in the total number of newborn hNSCs compared to either agent alone. Taken together, the *in vitro* and *in vivo* data showed that buspirone in combination with melatonin had a profile different from either agent alone.

This was a small exploratory study and the results must be interpreted with caution. Some reports suggest that high doses of buspirone above 40 mg may have antidepressant properties (Schweizer et al., 1986; Robinson et al., 1990). We found that melatonin-SR 3 mg enhanced the putative antidepressant properties of buspirone so that *low* dose buspirone (only 15 mg) taken in combination with melatonin yielded significant clinical improvement in MDD. In conjunction with the antidepressant efficacy, we found that the combination treatment had low adverse event liability. These findings have obvious clinical implications for the large number of subjects with MDD who do not respond adequately to currently available antidepressant medications. However, additional, larger studies evaluating combination buspirone and melatonin in MDD are needed to confirm these preliminary findings.

Contributors

Each listed author contributed to this paper.

Maurizio Fava, Steven D. Targum, Andrew A. Nierenberg, and Pamela C. Wedel contributed equally to the design, analysis, and execution of the clinical trial. Carrolee Barlow, Leo S. Bleicher, Todd A. Carter, René Hen, and Fred H. Gage contributed to the pre-clinical discovery platform, pre-clinical assays, study design and data analysis.

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Conflict of interest

Maurizio Fava, MD — Disclosures.

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Dr. Nierenberg is a full time employee of the Massachusetts General Hospital (MGH) and has disclosed all external sources of revenue to Harvard Medical School and Partners Health care to the best of his knowledge and in accordance with current regulations. External activities were limited to no more than 8 h per week. In the past 36 months (as of July 6, 2010) he has served as a consultant to: Appliance Computing Inc. (Mindsite), Brain Cells, Inc., Brandeis University, Bristol Myers Squibb, Clintara, Dianippon Sumitomo (Now Sunovion), Eli Lilly and Company, EpiQ, Novartis, PamLabs, PGx Health, Shire, Schering-Plough, Takeda Pharmaceuticals, and Targacept. He has consulted through the MGH Clinical Trials Network and Institute (CTNI): Astra Zeneca, Brain Cells, Inc., Dianippon Sumitomo/Sepracor, Johnson and Johnson, Labopharm, Merck, Methylation Science, Novartis, PGx Health, Shire, Schering-Plough, Targacept, and Takeda/Lundbeck Pharmaceuticals. He has received grant/research support through MGH from AHRQ, Cephalon, NIMH, PamLabs, Pfizer Pharmaceuticals, and Shire. Dr. Nierenberg received honoraria or travel expenses including CME activities from: APSARD, Belvoir Publishing, University of Texas Southwestern Dallas, Hillside Hospital, American Drug Utilization Review, American Society for Clinical Psychopharmacology, Bayamon Region Psychiatric Society, San Juan, PR, Baystate Medical Center, Canadian Psychiatric Association, Columbia University, Douglas Hospital/McGill University, IMEDEX, International Society for Bipolar Disorders, Israel Society for Biological Psychiatry, John Hopkins University, MJ Consulting, New York State, Massachusetts Association of College Counselors, Medscape, MBL Publishing, Physicians Postgraduate Press, Slack Publishing, SUNY Buffalo, University of Florida, University of Miami, University of Wisconsin, University of Pisa, and SciMed. Dr. Nierenberg is a presenter for the Massachusetts General Hospital Psychiatry Academy (MGHPA). The education programs conducted by the MGHPA were supported through Independent Medical Education (IME) grants from the following pharmaceutical companies in 2008: Astra Zeneca, Eli Lilly, and Janssen Pharmaceuticals; in 2009 Astra Zeneca, Eli Lilly, and Bristol-Myers Squibb. No speaker bureaus or boards since 2003. Dr. Nierenberg owns stock options in Appliance Computing, Inc. and Brain Cells, Inc. Additional income is possible from Infomedic.com depending on overall revenues of the company but no revenue has been received to date. Through MGH, Dr. Nierenberg is named for copyrights to: the Clinical Positive Affect Scale and the MGH Structured Clinical Interview for the Montgomery Asberg Depression Scale exclusively licensed to the MGH Clinical Trials Network and Institute (CTNI).

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Appendix A. Supplementary data

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References

- Adeagbo AS, Kadavil EA, Yousif M, Oriowo MA. Buspirone, a 5-HT(1A) receptor agonist, dilates the perfused rat uterine vascular bed through alpha(1)-adrenoceptor blockade. General Pharmacology 2000;34(5):357—62.
- Aimone JB, Deng W, Gage FH. Adult neurogenesis: integrating theories and separating functions. Trends in Cognitive Science 2010;14:325–37.
- Aimone JB, Deng W, Gage FH. Resolving new memories: a critical look at the dentate gyrus, adult neurogenesis, and pattern separation. Neuron 2011;70(4): 589–96.
- Alpert JE, Franznick DA, Hollander SB, Fava M. Gepirone extended-release treatment of anxious depression: evidence from a retrospective subgroup analysis in patients with major depressive disorder. Journal of Clinical Psychiatry 2004; 65(8):1069–75.
- Amsterdam JD, Brunswick DJ, Gibertini M. Sustained efficacy of gepirone-IR in major depressive disorder: a double-blind placebo substitution trial. Journal of Psychiatric Research 2004;38(3):259–65.
- Appelberg BG, Syvalahti EK, Koskinen TE, Mehtonen OP, Muhonen TT, Naukkarinen HH. Patients with severe depression may benefit from buspirone augmentation of selective serotonin reuptake inhibitors: results from a placebo-controlled, randomized, double-blind, placebo wash-in study. Journal of Clinical Psychiatry 2001;62(6):448–52.
- Bielski RJ, Cunningham L, Horrigan JP, Londborg PD, Smith WT, Weiss K. Gepirone extended-release in the treatment of adult outpatients with major depressive disorder: a double-blind, randomized, placebo-controlled, parallel-group study. Journal of Clinical Psychiatry 2008;69(4):571–7.
- Blier P, Bergeron R, De Montigny C. Selective activation of postsynaptic 5-HT1A receptors induces rapid antidepressant response. Neuropsychopharmacology 1997;16(5):333–8.
- Blier P, Ward NM. Is there a role for 5-HT1A agonists in the treatment of depression? Biological Psychiatry 2003;53(3):193–203.
- Boldrini M, Underwood MD, Hen R, Rosoklija GB, Dwork AJ, John Mann J, et al. Antidepressants increase neural progenitor cells in the human hippocampus. Neuropsychopharmacol 2009;34(11):2376–89.
- Bosker FJ, Westerink BH, Cremers TI, Gerrits M, van der Hart MG, Kuipers SD, et al. Future antidepressants: what is in the pipeline and what is missing? CNS Drugs 2004;18(11):705–32.
- Brown JP, Couillard-Despres S, Cooper-Kuhn CM, Winkler J, Aigner L, Kuhn HG. Transient expression of doublecortin during adult neurogenesis. Journal of Comprehensive Neurology 2003;467(1):1–10.
- Bourin M, Prica C. Melatonin receptor agonist agomelatine: a new drug for treating unipolar depression. Current Pharmaceutical Design 2009;15(14):1675–82.
- David DJ, Samuels BA, Rainer Q, Wang JW, Marsteller D, Mendez I, et al. Neurogenesis-dependent and -independent effects of fluoxetine in an animal model of anxiety/depression. Neuron 2009;62(4):479–93.
- Deakin JFW. A review of clinical efficacy of 5HT 1A agonists in anxiety and depression. Journal of Psychopharmacology 1993;7:283–9.
- Dranovsky A, Hen R. Hippocampal neurogenesis: regulation by stress and antidepressants. Biological Psychiatry 2006;59(12):1136–43.
- Dranovsky A, Picchini AM, Moadel T, Sisti AC, Yamada A, Kimura S, et al. Experience dictates stem cell fate in the adult hippocampus. Neuron 2011;70(5):908–23.
- Drevets WC. Neuroimaging and neuropathological studies of depression: implications for the cognitive-emotional features of mood disorders. Current Opinions in Neurobiology 2001;11(2):240–9.
- Duman RS, Voleti B. Signaling pathways underlying the pathophysiology and treatment of depression: novel mechanisms for rapid-acting agents. Trends in Neuroscience 2012;35(1):47–56.
- Eser D, Baghai TC, Möller HJ. Evidence of agomelatine's antidepressant efficacy: the key points. International Clinical Psychopharmacology 2007;22(Suppl. 2):S15—9.
- Eriksson PS, Perfilieva E, Bjork-Eriksson ET, Alborn AM, Nordborg C, Peterson DA, et al. Neurogenesis in the adult human hippocampus. Nature Medicine 1998; 4(11):1313–7.
- Fava M, Rush AJ. Current status of augmentation and combination treatments for major depressive disorder: a literature review and a proposal for a novel approach to improve practice. Psychotherapy and Psychosomatics 2006;75(3): 139–53.

- Gass P, Henn FA. Is there a role for neurogenesis in depression? Biological Psychiatry 2009;66(1):3-4.
- Gold PW, Chrousos GP. Organization of the stress system and its dysregulation in melancholic and atypical depression: high vs low CRH/NE states. Molecular Psychiatry 2002;7(3):254–75.
- Guy W. ECDEU assessment manual for psychopharmacology (revised). Rockville MD: Us Department of Health Education and Welfare, Public Health Service, Alcohol, Drug Abuse and Mental Health Administration: National Institute of Health, Psychopharmacology Research Branch; 1976.
- Hamilton MA. The assessment of anxiety states by rating. British Journal of Medical Psychology 1959;32(1):50–5.
- Hanson ND, Owens MJ, Nemeroff CB. Depression, antidepressants, and neurogenesis: a critical appraisal. Neuropsychopharmacol 2011;36(13):2589–602.
- Heiser J, Wilcox CS. Serotonin 5HT 1A receptor agonists as antidepressants; pharmacological rationale and evidence for efficacy. CNS Drugs 1998;10:343–53.
- Henn FA, Vollmayr B. Neurogenesis and depression: etiology or epiphenomenon? Biological Psychiatry 2004;56(3):146–50. Holick KA, Lee DC, Hen R, Dulawa SC. Behavioral effects of chronic fluoxetine in
- Holick KA, Lee DC, Hen R, Dulawa SC. Behavioral effects of chronic fluoxetine in BALB/cJ mice do not require adult hippocampal neurogenesis or the serotonin 1A receptor. Neuropsychopharmacology 2008;33:406–17.
- Jacobs BL, Praag H, Gage FH. Adult brain neurogenesis and psychiatry: a novel theory of depression. Molecular Psychiatry May 2000;5(3):262–9.
- Kasper S, Hamon M. Beyond the monoaminergic hypothesis: agomelatine, a new antidepressant with an innovative mechanism of action. World Journal of Biological Psychiatry 2009;10(2):117–26.
- Keller MB, Ruwe FJ, Janssens CJ, Sitsen JM, Jokinen R, Janczewski J. Relapse prevention with gepirone ER in outpatients with major depression. Journal of Clinical Psychopharmacology 2005;25(1):79–84.
- Landen M, Bjorling G, Agren H, Fahlen T. A randomized, double-blind, placebocontrolled trial of buspirone in combination with an SSRI in patients with treatment-refractory depression. Journal of Clinical Psychiatry 1998;59(12):664–8.
- Lapierre YD, Silverstone P, Reesal RT, Saxena B, Turner P, Bakish D, et al. A Canadian multicenter study of three fixed doses of controlled-release ipsapirone in outpatients with moderate to severe major depression. Journal of Clinical Psychopharmacology 1998;18(4):268–73.
- Laughren TP, Temple RJ, Unger EF, Bhattaram A, Dinh PV, Fossom L, et al. Vilazodone: clinical basis for the US food and drug administration's approval of a new antidepressant. Journal of Clinical Psychiatry 2011;72(9):1166–73.
- Li Y, Luikart BW, Birnbaum S, Chen J, Kwon CH, Kernie SG, et al. TrkB regulates hippocampal neurogenesis and governs sensitivity to antidepressive treatment. Neuron 2008;59(3):399–412.
- Lieberman JA, Greenhouse J, Hamer RM, Krishnan KR, Nemeroff CB, Sheehan DV. Comparing the effects of antidepressants: consensus guidelines for evaluating quantitative review of antidepressant efficacy. Neuropsychopharmacology 2005; 30:445–60.
- Lucassen PJ, Meerlo P, Naylor AS, van Dam AM, Dayer AG, Fuchs E, et al. Regulation of adult neurogenesis by stress, sleep disruption, exercise and inflammation: implications for depression and antidepressant action. European Neuropsychopharmacology 2010;20:1–17.
- Malberg JE, Eisch AJ, Nestler EJ, Duman RS. Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. Journal of Neuroscience 2000;20(24):9104–10.
- Miller BH, Schultz LE, Gulati A, Cameron MD, Pletcher MT. Genetic regulation of behavioral and neuronal responses to fluoxetine. Neuropsychopharmacol 2008; 33:1312–22.
- Nassir Ghaemi S. Why antidepressants are not antidepressants: STEP-BD, STAR*D, and the return of neurotic depression. Bipolar Disorders 2008;10(8):957–68.
- Ramirez-Rodriguez G, Klempin F, Babu H, Benitez-King G, Kempermann G. Melatonin modulates cell survival of new neurons in the hippocampus of adult mice. Neuropsychopharmacology 2009;34(9):2180–9.
- Rickels KJ, Amsterdam JD, Clary C, Hassman J, London J, Puzzuoli G, et al. Buspirone in depressed outpatients: a controlled study. Psychopharmacology Bulletin 1990;26(2):163–7.
- Rickels K, Amsterdam JD, Clary C, Puzzuoli G, Schweizer E. Buspirone in major depression: a controlled study. Journal of Clinical Psychiatry 1991;52(1):34–8.
- Rickels K, Khalid-Khan S, Rynn M. Buspirone in the treatment of anxiety disorders. In: Nutt DJ, Ballenger JC, editors. Anxiety disorders. Blackwell Publishing; 2003. p. 381–97.
- Rickels K, Athanasiou M, Robinson DS, Gibertini M, Whalen H, Reed CR. Evidence for efficacy and tolerability of vilazodone in the treatment of major depressive disorder: a randomized, double-blind, placebo-controlled trial. Journal of Clinical Psychiatry 2009;70(3):326–33.
- Robinson DS, Rickels K, Feighner J, Fabre Jr LF, Gammans RE, Shrotriya RC, et al. Clinical effects of the 5-HT1A partial agonists in depression: a composite analysis of buspirone in the treatment of depression. Journal of Clinical Psychopharmacology 1990;10(3 Suppl.):675–76S.
- Rush AJ. STAR*D: what have we learned? American Journal of Psychiatry 2007; 164(2):201-4.
- Rush AJ, Gullion CM, Basco MR, Jarrett RB, Trivedi MH. The Inventory of depressive symptomatology (IDS): psychometric properties. Psychological Medicine 1996; 26(3):477–86.
- Rush AJ, Trivedi MH, Ibrahim HM, Carmody T, Arnow B, Klein DN, et al. The 16-item quick inventory of depressive symptomatology (QIDS), clinician rating (QIDS-C), and self-report (QIDS-SR): a psychometric evaluation in patients with chronic major depression. Biological Psychiatry 2003;54(5):573–83.

- Rush AJ, Trivedi MH, Wisniewski SR, Nierenberg AA, Stewart JW, Warden D, et al. Acute and longer-term outcomes in depressed outpatients requiring one or several treatment steps: a STAR*D report. American Journal of Psychiatry 2006;163(11):1905–17.
- Rush AJ, Wisniewski SR, Warden D, Luther JF, Davis LL, Fava M, et al. Selecting among second-step antidepressant medication monotherapies. Archives of General Psychiatry 2008;65(8):870–81.
- Sahay A, Hen R. Adult hippocampal neurogenesis in depression. Nature Neuroscience 2007;10(9):1110–5.
- Santarelli LM, Saxe C, Gross A, Surget F, Battaglia S, Dulawa N, et al. Requirement of hippocampal neurogenesis for the behavioral effects of antidepressants. Science 2003;301(5634):805–9.
- Schweizer EE, Amsterdam J, Rickels K, Kaplan M, Droba M. Open trial of buspirone in the treatment of major depressive disorder. Psychopharmacology Bulletin 1986;22(1):183-5.
- Sheehan DV, Lecrubier Y, Sheehan KH, Amorim P, Janavs J, Weiller E, et al. The mininternational neuropsychiatric interview (M.I.N.I.): the development and validation of a structured diagnostic psychiatric interview for DSM-IV and ICD-10. Journal of Clinical Psychiatry 1998;59(Suppl. 20):22–33. guiz 34–57.
- Sheline YI, Gado MH, Kraemer HC. Untreated depression and hippocampal volume loss. American Journal of Psychiatry 2003:160(8):1516—8.
- Snyder JS, Choe JS, Clifford MA, Jeurling SI, Hurley P, Brown A, et al. Adult-born hippocampal neurons are more numerous, faster maturing, and more involved in behavior in rats than in mice. Journal of Neuroscience 2009; 29(46):14484–95.
- Snyder JS, Soumier A, Brewer M, Pickel J, Cameron HA. Adult hippocampal neurogenesis buffers stress responses and depressive behaviour. Nature 2011;476: 458–61.
- Soumier A, Banasr M, Lortet S, Masmejean F, Bernard N, Kerkerian-Le Goff L, et al. Mechanisms contributing to the phase-dependent regulation of neurogenesis by the novel antidepressant, agomelatine, in the adult hippocampus. Neuropsychopharmacology 2009;34:2390–403.

- Sramek JJ, Tansman M, Suri A, Hornig-Rohan M, Amsterdam JD, Stahl SM, et al. Efficacy of buspirone in generalized anxiety disorder with coexisting mild depressive symptoms. Journal of Clinical Psychiatry 1996;57(7):287–91.
- Stahl SM, Kaiser L, Roeschen J, Keppel Hesselink JM, Orazem J. Effectiveness of ipsapirone, a 5-HT-1A partial agonist, in major depressive disorder: support for the role of 5-HT-1A receptors in the mechanism of action of serotonergic antidepressants. International Journal of Neuropsychopharmacology 1998;1(1):11-8.
- Surget A, Saxe M, Leman S, Ibarguen-Vargas Y, Chalon S, Griebel G, et al. Drugdependent requirement of hippocampal neurogenesis in a model of depression and of antidepressant reversal. Biological Psychiatry 2008;64:293—301.
- Svendsen CN, ter Borg MG, Armstrong RJ, Rosser AE, Chandran S, Ostenfeld T, et al. A new method for the rapid and long term growth of human neural precursor cells. Journal of Neuroscience Methods 1998;85(2):141–52.
- Targum SD, Busner J, Young AH. Targeted scoring criteria reduces variance in global impressions. Human Psychopharmacology: Clinical and Experimental 2008;23: 628–33.
- Thase ME, Howland RH, Friedman ES. Treating antidepressant nonresponders with augmentation strategies: an overview. Journal of Clinical Psychiatry 1998;59-(Suppl. 5):5–12. discussion 13–5.
- Trivedi MH, Fava M, Wisniewski SR, Thase ME, Quitkin F, Warden D, et al. Medication augmentation after the failure of SSRIs for depression. New England Journal of Medicine 2006;354(12):1243–52.
- van Praag H, Schinder AF, Christie BR, Toni N, Palmer TD, Gage FH. Functional neurogenesis in the adult hippocampus. Nature 2002;415(6875):1030–4.
- Wang JW, David DJ, Monckton JE, Battaglia F, Hen R. Chronic fluoxetine stimulates maturation and synaptic plasticity of adult-born hippocampal granule cells. Journal of Neuroscience 2008;28(6):1374–84.
- Warner-Schmidt JL, Duman RS. Hippocampal neurogenesis: opposing effects of stress and antidepressant treatment. Hippocampus 2006;16(3):239–49.
- Wilcox CS, Ferguson JM, Dale J, Heiser JF. A double-blind trial of low- and high-dose ranges of gepirone-ER compared with placebo in the treatment of depressed outpatients. Psychopharmacol Bulletin 1996;32(3):335–42.