JAMA Psychiatry | Original Investigation

Association Between Population Density and Genetic Risk for Schizophrenia

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IMPORTANCE Urban life has been proposed as an environmental risk factor accounting for the increased prevalence of schizophrenia in urban areas. An alternative hypothesis is that individuals with increased genetic risk tend to live in urban/dense areas.

OBJECTIVE To assess whether adults with higher genetic risk for schizophrenia have an increased probability to live in more populated areas than those with lower risk.

DESIGN, SETTING, AND PARTICIPANTS Four large, cross-sectional samples of genotyped individuals of European ancestry older than 18 years with known addresses in Australia, the United Kingdom, and the Netherlands were included in the analysis. Data were based on the postcode of residence at the time of last contact with the participants. Community-based samples who took part in studies conducted by the Queensland Institute for Medical Research Berghofer Medical Research Institute (QIMR), UK Biobank (UKB), Netherlands Twin Register (NTR), or QSkin Sun and Health Study (QSKIN) were included. Genome-wide association analysis and mendelian randomization (MR) were included. The study was conducted between 2016 and 2018.

EXPOSURES Polygenic risk scores for schizophrenia derived from genetic data (genetic risk is independently measured from the occurrence of the disease). Socioeconomic status of the area was included as a moderator in some of the models.

MAIN OUTCOMES AND MEASURES Population density of the place of residence of the participants determined from census data. Remoteness and socioeconomic status of the area were also tested.

RESULTS The QIMR participants (15 544; 10 197 [65.6%] women; mean [SD] age, 54.4 [13.2] years) living in more densely populated areas (people per square kilometer) had a higher genetic loading for schizophrenia ($r^2 = 0.12\%$; $P = 5.69 \times 10^{-5}$), a result that was replicated across all 3 other cohorts (UKB: 345 246; 187 469 [54.3%] women; age, 65.7 [8.0] years; NTR: 11 212; 6727 [60.0%] women; age, 48.6 [17.5] years; and QSKIN: 15 726; 8602 [54.7%] women; age, 57.0 [7.9] years). This genetic association could account for 1.7% (95% CI, 0.8%-3.2%) of the schizophrenia risk. Estimates from MR analyses performed in the UKB sample were significant (b = 0.049; $P = 3.7 \times 10^{-7}$ using GSMR), suggesting that the genetic liability to schizophrenia may have a causal association with the tendency to live in urbanized locations.

CONCLUSIONS AND RELEVANCE The results of this study appear to support the hypothesis that individuals with increased genetic risk tend to live in urban/dense areas and suggest the need to refine the social stress model for schizophrenia by including genetics as well as possible gene-environment interactions.

JAMA Psychiatry. doi:10.1001/jamapsychiatry.2018.1581 Published online June 23, 2018. **Editorial**

Supplemental content

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Corresponding Author: Lucía Colodro-Conde, PhD, QIMR Berghofer Medical Research Institute, Locked Bag 2000, Royal Brisbane Hospital QLD 4029, Australia (lucia.colodroconde@qimrberghofer .edu.au). n 2011, Lederbogen and colleagues¹ published a functional magnetic resonance imaging study that showed greater brain activation of the stress-processing pathways in participants living in urban vs rural areas and suggested there and in a later study² that the greater social stress of urban living could explain the well-documented higher prevalence of schizophrenia observed in urban than rural environments (odds ratio, 1.72; 95% CI, 1.53-1.92).³ Herein, we investigate an alternative, but not incompatible, explanation: people with higher genetic risk for schizophrenia tend to live in more urbanized areas owing to selective migration⁴ in either past or current generations.

eAppendix 1 in the Supplement presents a short review of the literature in this area. In summary, living in an urban environment, which is itself partially heritable,⁵ is associated with increased risk of developing schizophrenia after controlling for potential confounders (age, sex, ethnicity, drug use, social class, family history, and season of birth) and using different measures of urbanicity (population size or density^{6,7}), window of exposure (birth, 4,8 upbringing, 1,6 or illness onset 7,9), and disease definition (narrow schizophrenia or broad psychosis^{7,10}). Although the association is established, its putative (familial) environmental or genetic components are unclear. It has been suggested that approximately 30% of all schizophrenia cases could be potentially prevented if the exposure to urban environments was removed, assuming urban environment is a causal factor.10 However, it is not clear whether urban residence has a causal effect on mental health or whether urban residence is a consequence of the disease (eg, migration to the city of people in the prodromal stages of the disorder⁶).

In the present study, we sought to examine the nature of this association by testing whether adults older than 18 years with higher genetic risk for schizophrenia are more likely to live in urbanized and populated areas (measured as population density) than those with lower genetic risk. If so, this finding would suggest that the higher prevalence of schizophrenia in cities is not only a consequence of the urban environment. This determination has been made possible by the advances in the identification of common genetic variants associated with schizophrenia in large discovery samples¹¹ and the development of polygenic risk scores (PRS) in independent samples.¹² In addition, we checked that the association could not be explained by differences in socioeconomic status (SES) of the residential areas. We also investigated the direction of causation between schizophrenia and population density using multi-instrument mendelian randomization (MR). 13,14 For completeness, we present the estimates of the twin heritability and genome-wide association analyses (GWAS) of our main phenotypes.

Methods

Cohorts and Variables

We performed the analyses using a discovery cohort of 15 544 participants genotyped as part of a series of studies of general health conditions conducted by the Genetic Epidemiology Unit at Queensland Institute for Medical Research Berghofer Medi-

Key Points

Question Are adults with higher genetic risk for schizophrenia more likely to live in urbanized and populated areas than those with lower risk?

Findings In this cross-sectional study of 4 community-based samples from Australia, the United Kingdom, and the Netherlands (N = 504 130), significantly higher genetic loading for schizophrenia was identified in participants living in more densely populated areas; mendelian randomization on a subsample suggests that schizophrenia may have a causal association with the tendency to live in urban areas.

Meaning The higher rates of schizophrenia in cities may be accentuated by selective migration to cities of participants with higher genetic risks.

cal Research Institute (QIMR), Australia^{15,16} (**Table 1**; eAppendix 2, eTable 1, and eFigure 1 in the Supplement provide cohort and variable descriptions). We used the UK Biobank (UKB) ($n = 456\ 426$), ^{17,18} the Netherlands Twin Register (NTR) ($n = 16\ 434$), ¹⁹ and the Australian QSkin Sun and Health Study (QSKIN) sample ($n = 15\ 726$) ²⁰ to replicate and extend our analyses (Table 1; eAppendixes 3-5, eTable 2, eTable 3, eFigures 2-5 in the Supplement provide cohort and variable descriptions).

The QIMR Berghofer Medical Research Institute-Human Research Ethics Committee approved the study. The study was conducted between 2016 and 2018.

Statistical Analysis

Variance Component Analysis

We analyzed data from 5894 twins from the QIMR sample to estimate the contribution of additive genetic influences (narrow sense heritability), shared/familial environment, and unique environment to the interpersonal differences in population density, remoteness, and SES of the residential area (eAppendix 6 in the Supplement provides more information on twin and family studies). We used the OpenMx²⁴ package in R²⁵ to estimate the parameters of the mixed models. Significance of the variance components was tested using likelihood ratio tests on nested models. ²⁶ We used the same approach to replicate the results on population density in the NTR cohort.

In the QIMR data, we fit a gene-environment moderator effect model²⁷ that allows the variance components (heritability, shared environment, and unique environment) to vary across age (eAppendix 7 in the Supplement). This approach reflected previous results from Whitfield et al⁵ that suggested that heritability of rural/urban living in Australia increases with age. Finally, we estimated the genetic and environmental correlations between population density, remoteness, and SES using bivariate twin models.²⁶ All models included age, age,² sex, age × sex, age² × sex, GWAS array, and 4 genetic principal components as covariates.

PRS Analysis

The PRS in the QIMR sample were calculated from the imputed genotype dosage scores using GWAS summary statistics from the GWAS meta-analysis from the 2014 Psychiatric

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Table 1. Description	of the Colloits Used	a for the Analyses

	Discovery Cohort	Replication Cohorts						
Variable	QIMR ^{15,16} (Australia)	UKB ^{17,18} (United Kingdom)	NTR ¹⁹ (the Netherlands)	QSKIN ²⁰ (Australia [limited to Queensland]				
Sample size, No.	15 544	456 426 (345 246 with PRS)	16 434 (11 212 with PRS)	15 726				
Inclusion criteria ^b	Adult genotyped participants of QIMR studies	Genotyped participants of the UKB	Adult genotyped participants of the NTR	Unrelated (GRM <0.1) adult genotyped participants of the QSKIN who had not previously participated in QIMR studies				
Demographics	10 197 (65.6%) Women	187 469 (54.3%) Women	6727 (60.0%) Women	8602 (54.7%) Women				
	Age, mean (SD): 54.4 (13.2)	Age, mean (SD): 65.7 (8.0)	Age, mean (SD): 48.6 (17.5)	Age, mean (SD), 57.0 (7.9)				
	From 7015 families		From 4456 families					
	Sample comprises 1119 complete MZ pairs of twins, 1104 complete DZ pairs, and 1448 singleton twins	Sample comprises 345 258 unrelated individuals (GRM <0.05)	Sample comprises 1740 complete MZ pairs, 1114 DZ complete twin pairs, and 812 singleton twins					
Genetic data	Participants genotyped using commercial arrays	Participants genotyped using 2 closely related arrays (the UK BiLEVE and the UK Biobank Axiom Arrays)	Participants genotyped using commercial arrays	Same as QIMR				
	Genotype data cleaned (by batch) for call rate (≥95%)	We used HRC imputed data provided by the biobank	Genotype data cleaned for call rate (≥95%)					
	MAF (≥1%)	Variants with MAF <0.005%, missingness <0.05, pHWE <10 ⁻⁶ , and MAC >5 excluded	MAF ≥0.5%					
	Hardy-Weinberg equilibrium ($P \ge 10^{-3}$), GenCall score (≥0.15 per genotype; mean, ≥0.7), standard Illumina filters	Data were checked for non-European ancestry	Hardy-Weinberg equilibrium $(P \ge 10^{-12})$, allele frequency difference with GONL <0.10					
	Data checked for pedigree, sex, and mendelian errors and for non-European ancestry		Data checked for pedigree, sex, and mendelian errors and for non-European ancestry ²¹					
	Imputation to the 1000 genomes (phase 3, release 5) performed on the Michigan Imputation Server ²²							
Measures of population density and SES	Population density, remoteness, and SES variables generated from the postcode provided by participants at the time of last contact (1990-2015)	From the Easting and Northing coordinates rounded to the kilometer, we performed reverse geocoding to identify the postcode district in which the participants likely lived	Population density and SES derived from the most recent participants' postcodes	Population density and SES variables were generated from the postcode provided by the participants at the time of last contact (2010-2012)				
	We matched postcodes to the latest census data collected by the Australian Bureau of Statistics (2016 for population density, 2011 for remoteness and SES)	We crossed this information with the population density by postcode district calculated in the 2011 census	Numbers corresponded to the neighborhood data published in 2015-2016 by the Netherlands's national statistical agency (CBS), which defines a neighborhood as the part of a municipality that is homogeneously demarcated from either a demographic or socioeconomic structure	We matched the postcodes as described in the QIMR sample				
	Population density expressed in number of residents per squared kilometer	Population density expressed in number of residents per hectare	Population density expressed in number of residents per squared kilometer	Population density expressed in number of residents per squared kilometer				
	SES based on the IRSAD, 23 which can be used to measure socioeconomic well-being in a continuum, from the most disadvantaged areas (low values) to the most advantaged areas (high values)	We used the Townsend deprivation index ^{17,18} as a measure of SES	SES measured using the average personal income per person and the average market value of residential properties	SES based on the IRSAD, ²³ as described in the QIMR sample				
	Mean (SD) population density was 1169 (1350) people/km ² (range, 0.01-5506)	Mean (SD) population density was 24.10 (25.1) inhabitants per hectare (ie, 2410 per km²) (range, 0.1-222.5)	Mean (SD) population density in the total sample was 4669 (3793) (range, 1-17797)	Mean (SD) population density was 748 people/km² (945) (range, 0.01-3771)				

Abbreviations: DZ, dizygotic; GONL, Genome of the Netherlands; GRM, genetic relationship matrices; HRC, Haplotype Reference Consortium; IRSAD, Index of Relative Socioeconomic Advantage and Disadvantage; MAC, minor allele count; MAF, minor allele frequency; MZ, monozygotic; NTR, Netherlands Twin Register; PHWE, P value for the Hardy-Weinberg test statistic; PRS, polygenic risk scores; QIMR, QIMR Berghofer Medical Research Institute; QSKIN, QSkin

Sun and Health Study; SES, socioeconomic status; UKB, United Kingdom Biobank.

^a The Supplement contains documents on genetic and phenotypic data for all cohorts.

 $^{^{\}rm b}$ European ancestry was an inclusion criterion across all cohorts.

Genomics Consortium Schizophrenia Working Group (36 989 cases and 113 075 controls)11 following the method described by Wray et al. 12 We excluded single-nucleotide polymorphisms (SNPs) with low imputation quality (r^2 <0.6) or minor allele frequency (MAF) below 1%. We selected the most significant independent SNPs using PLINK1.9²⁸ to correct for signal redundancy owing to linkage disequilibrium (LD) (criteria LD r^2 <0.1 within windows of 10 mb). We calculated 8 different PRS using different P value thresholding of the GWAS summary statistics (eTable 1 in the Supplement provides the number of SNPs included for each threshold, and eFigure 1 in the Supplement shows histograms of PRS for schizophrenia). We used mixed models to test the association between neighborhood variables (population density, remoteness, and SES) and PRS and set the significance threshold to 3.15×10^{-3} to account for multiple testing (eAppendix 8 in the Supplement).

We replicated the PRS analysis for the neighborhood variables in the UKB, NTR, and QSKIN samples using the same GWAS summary statistics¹¹ and mixed model approach (eAppendix 8 in the Supplement). eAppendixes 3-5 in the Supplement provide differences in phenotypic and genetic data and PRS calculation between these samples.

Genome-wide Association Analyses

We performed GWAS of population density or SES in our largest sample, the UKB, using BOLT-LMM²⁹ including age, age, sex, age \times sex, age² \times sex, and 4 genetic principal components ³⁰ on 12 272 635 SNPs with MAF greater than 0.005%. We ran additional GWAS of population density controlling for SES and of SES controlling for population density.

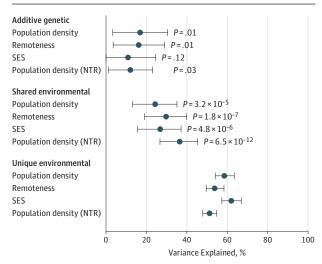
We also conducted GWAS of population density in the QIMR, NTR, and QSKIN cohorts. In the QIMR sample, we used RAREMETALWORKER³¹ and controlled for age, age,² sex, age × sex, age² × sex, SES, GWAS array, and 4 genetic principal components as covariates. We explicitly corrected for relatedness using the kin pedigree option. Single-nucleotide polymorphisms with MAF less than 0.5% or imputation r^2 <0.6 were excluded, leaving 8 495 074 SNPs for analyses. In the NTR, the GWAS, corrected for age and sex, was performed using GCTA-MLMA^{32,33} to account for relatedness and population stratification using 2 genetic relationship matrices (one corrects for the related individuals; the second corrects for the population stratification in the distantly related individuals) and 5 genetic principal components. The number of markers included after quality control (r^2 >0.8 and MAF<0.01%) was 7 636 917. Finally, we performed the GWAS in QSKIN using PLINK, version 1.90b4.128 with age, sex, and 4 genetic principal components as covariates, using a total of 7 672 045 markers after selecting those with $r^2>0.6$ and MAF less than 0.01%.

We used LD score regressions³⁴ to confirm the SNP heritability and the genetic correlations between the measures of population density across samples. eAppendixes 2-5 in the Supplement provide details on genetic data of the 4 cohorts.

Mendelian Randomization

Mendelian randomization methods allow us to generate hypotheses concerning the direction of causation between 2 heritable variables.¹³ Herein, we relied on the 2014 GWAS

Figure 1. Percentage of Variance



Additive genetic, shared environmental, and unique environmental factors in population density, remoteness, and socioeconomic status (SES) in the Queensland Institute of Medical Research cohort (discovery). Results for population density in the Netherlands Twin Register (NTR) cohort are presented for replication. *P* values are not applicable for unique environmental factors.

meta-analysis summary results from the Psychiatric Genomics Consortium for schizophrenia and the GWAS results calculated from our samples for population density and SES. We used MR-Base³⁵ (TwoSampleMR R package³⁶) and GSMR³⁷ to conduct MR using known schizophrenia SNPs as instruments, thus testing the selection hypothesis that having a higher propensity to schizophrenia (ie, higher PRS) may have a possible causal association with the tendency to live in a denser and less remote area (eAppendix 9 in the Supplement gives more details on MR and our analysis). We also investigated the reverse hypothesis (population density or SES inducing onset of schizophrenia) using GWAS results from our largest sample, the UKB.

Results

Variance Component Analysis

Population density, remoteness, and SES were all significantly correlated at a phenotypic level in all samples considered (eAppendix 10 and eTable 4 in the Supplement). Population density and remoteness were heritable (heritability $[h^2]$) in the QIMR sample (Figure 1) (h^2 for population density = 16.9%; 95% CI, 3.4-30.4; P = .01; h^2 for remoteness = 16.3%; 95% CI, 3.5-29.0; P = .01); the heritability of SES was not significant ($h^2 = 11.0\%$; 95% CI, 0.00-24.6; P = .12). Shared environment (common environment $[c^2]$) effects explained a more substantial and highly significant proportion of the trait variance (Figure 1) (c^2 for population density = 24.3%; 95% CI, 13.1-35.1; c^2 for remoteness = 29.1%; 95% CI, 19.0-40.0; and c^2 for socioeconomic status = 26.8%; 95% CI, 15.6-37.1; all P < .001), which highlights that people tend to live with or close to their parents or other relatives. Population density was also heritable in the NTR (h^2 = 12.1%; 95%

CI, 1.3-23.2; P = .28) and showed shared environment sources of variance ($c^2 = 36.5$; 95% CI, 26.8-45.7; $P = 7 \times 10^{-12}$).

In the QIMR cohort, population density was more heritable and less influenced by shared environmental sources as participants became older (eAppendix 11 in the Supplement), with the heritability increasing from 9.0% to 25.6% between ages 20 and 80 years. Over the same lifespan, c^2 decreased from 44.5% to less than 10.0% (eAppendix 11 in the Supplement), but variance explained by unique environmental sources (including measurement error) remained constant (eAppendix 11 in the Supplement). Similar results were obtained using standardized estimates, suggesting constant phenotypic variance across age. In addition, population density, remoteness, and SES shared environmental influences as indicated by significant environmental correlations from the twin models (eTable 4 in the Supplement).

Polygenic Risk Scores Analysis

Figure 2 shows the percentage of variance of the population density of the place of residence explained by the PRS for schizophrenia, with and without controlling for SES. In the QIMR sample, PRS calculated from all semi-independent SNPs across the genome (Figure 2) explained the greatest amount of variance in population density ($r^2 = 0.12\%$; $P = 5.69 \times 10^{-5}$), and still explained ($r^2 = 0.074\%$; P < .001) when accounting for SES. Schizophrenia PRS also were significantly associated with remoteness ($r^2 = 0.06\%$; P = .003) when including all of the independent SNPs, although the association disappeared when correcting for SES. eFigure 6 and eAppendix 12 in the Supplement provide information on the association between genetic risk for schizophrenia, remoteness, and SES. We did not find evidence of interactions between sex or age and PRS for schizophrenia (P > .05) contributing to population density or remoteness in the QIMR sample.

The association between schizophrenia risk score and population density was replicated in the NTR (r^2 = 0.14%; P = 8.3 × 10⁻⁴ and r^2 = 0.073%; P = .002 when correcting for SES), in the UKB (r^2 = 0.088%; P = 7.7 × 10⁻⁵⁹; r^2 = 0.012%; P = 1.2 × 10⁻¹¹ when accounting for SES) and in QSKIN (r^2 = 0.027%; P = .02 and r^2 = 0.015%; P = .047 when correcting for SES) (Figure 2). All correlations were in the same direction, pointing to increased PRS for participants living in more densely populated areas. Results for remoteness and SES in QSKIN were consistent with those in QIMR and are presented in eFigure 7 in the Supplement.

In addition, we tested the association between SES and schizophrenia PRS. The association did not reach statistical significance in the QIMR or QSKIN sample when taking into account multiple testing (P > .01) (eFigures 6 and 7 in the Supplement) but was significant in the UKB ($r^2 = 0.084\%$; $P = 8.6 \times 10^{-64}$, correcting for population density) (eFigure 8 in the Supplement) likely because of the gain of power owing to its very large sample size.

Genome-wide Association Analyses

Six genomic regions reached genome-wide significance for population density in the UKB, and this number increased to 12 when correcting for SES. Similarly, we identified 13 loci as-

sociated with SES in the UKB when correcting for population density. We observed fewer significant associations with population density or remoteness in the smaller samples, but they did not correspond to the SNP associations found in the UKB. The SNP heritability ranged from 0.6% (QIMR: SE, 3.2%) to 9.3% (QSKIN: SE, 4.0%) as estimated by LD score regression (eTable 5 in the Supplement). The genetic correlation between population density across samples ranged from 0.30 (SE, 0.44; P = .49, QSKIN-NTR) to 0.61 (SE, 0.29; P = .04, UKB-NTR). eAppendix 13, eFigures 9-16, and eTable 5 in the Supplement provide all GWAS and LD score regression results (including GWAS meta-analysis).

Mendelian Randomization

Finally, we selected between 88 and 94 genome-wide significant SNPs for schizophrenia as instruments to perform MR analyses with population density as the outcome variable after excluding SNPs showing evidence of pleiotropic effects by the heterogeneity in dependent instruments outlier analysis (implemented in the GSMR software).³⁷ These numbers are consistent with the 108 independent associations reported by the Psychiatric Genomics Consortium¹¹; the difference arose from SNPs not being present or not passing quality control in GWAS of population density and SES.

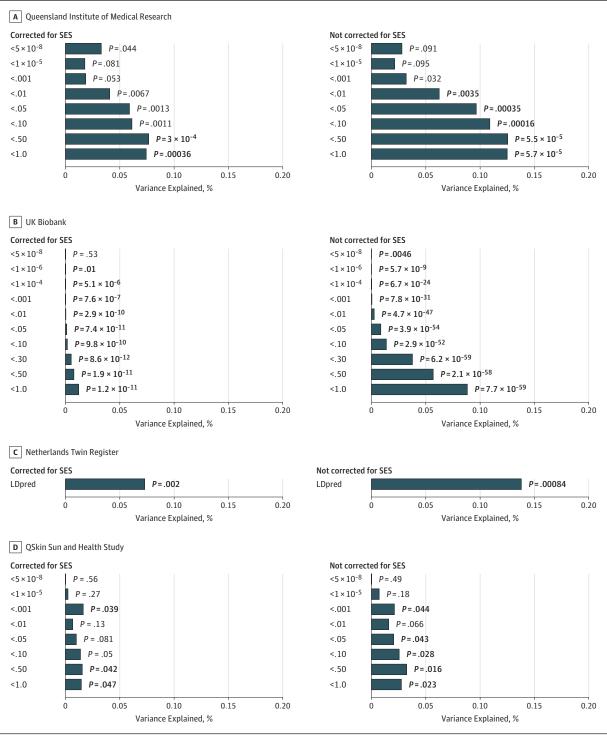
Estimates from MR analyses performed in the UKB sample were significant (b = 0.049; $P = 3.7 \times 10^{-7}$ using GSMR) (Table 2, Figure 3), suggesting that the genetic liability to schizophrenia has a causal association with the tendency to live in urbanized locations. We observed similar effect sizes in all other samples, although the MR results were not significant. We found no evidence of confounding heterogeneity of effect sizes (P > .30) or from pleiotropy (P > .05) using the tests implemented in MR-Base. Reverse MR testing (propensity to live in a more dense or low socioeconomic status area as a cause of schizophrenia) was only suggestive of a (larger) association with population density or SES on schizophrenia (b = 0.20; P = .01using GSMR) as it did not survive multiple testing correction. This lack of evidence could have been the result of reduced statistical power, as both genetic instruments had a smaller number of SNPs (n = 12) (Table 2; eAppendix 14 and eTables 6-10 in the Supplement provide detailed MR results).

Discussion

The present study investigated the association between genetic risk for schizophrenia and characteristics of a person's place of residence (population density, remoteness, and SES) to test the genetic nature of the association between schizophrenia and population density and infer the direction of causation. We used data on where people live collected as part of 4 studies from 3 countries (Australia, United Kingdom, and the Netherlands) for a total of 504 130 participants.

In all 4 nonclinical cohorts, genetic risk for schizophrenia was associated with greater population density of the postcode of residence beyond what could be explained by SES of the area (Figure 2). Results were consistent for remoteness. Our results show that the geographic distribution of the genetic risk for

Figure 2. Percentage of Variance of the Population Density of the Place of Residence Explained by the Genetic Risk for Schizophrenia, Both Without and With Socioeconomic Status (SES) as a Covariate



A-D, Genetic risk for schizophrenia showed positive effects for population density in all significant associations. Results in bold highlight significant results after correction for multiple testing. In the discovery Queensland Institute of Medical Research sample, we accounted for the number of tests performed. In the Netherlands Twin Register, UK Biobank, and QSkin Sun and Health Study,

we used a significance threshold of .05 as we aimed to replicate the results found with the polygenic risk scores (PRS) calculated over all independent genomic regions. However, when multiple PRS were available, we present all of the results for completeness. Python-based software (LDpred) was used to conduct the analyses in the NTR cohort.

schizophrenia is not uniform and that participants with higher genetic risk levels live in areas with higher population density over what is expected by chance. We also found a significant association between genetic risk for schizophrenia and SES of the

Table 2. Summary of the MR-Based Analysis of Schizophrenia and Population Density Across the 4 Cohorts

	QIMR			UKB		NTR			QSKIN			
Method	No. of SNPs	b (SE)	<i>P</i> Value	No. of SNPs	b (SE)	P Value	No. of SNPs	b (SE)	<i>P</i> Value	No. of SNPs	b (SE)	P Value
Fixed-effects meta-analysis (simple SE)	93	0.045 (0.050)	.36	92ª	0.054 (0.009) ^a	1.6 × 10 ^{-8a}	88	0.043 (0.050)	.37	94	0.021 (0.049)	.67
Fixed-effects meta-analysis (delta method)	93	0.047 (0.051)	.35	92 ^a	0.049 (0.01) ^a	3.4×10^{-7a}	88	0.042 (0.050)	.40	94	0.019 (0.050)	.69
Random-effects meta-analysis (delta method)	93	0.043 (0.054)	.43	92ª	0.046 (0.013) ^a	5.8 × 10 ^{-4a}	88	0.042 (0.050)	.40	94	0.019 (0.050)	.69
Maximum likelihood	93	0.045 (0.051)	.37	92 ^a	0.056 (0.01) ^a	9.6×10^{-9a}	88	0.044 (0.050)	.37	94	0.021 (0.050)	.66
MR Egger	93	0.49 (0.26)	.06	92	0.16 (0.06)	.013	88	-0.006 (0.240)	.97	94	0.27 (0.231)	.24
Weighted median	93	0.11 (0.075)	.13	92ª	0.053 (0.015) ^a	7.7 × 10 ^{-4a}	88	0.074 (0.074)	.31	94	0.026 (0.072)	.71
Inverse variance weighted	93	0.045 (0.056)	.41	92ª	0.053 (0.013) ^a	1.0 × 10 ^{-4a}	88	0.043 (0.050)	.37	94	0.021 (0.049)	.67
GSMR	93	0.039 (0.051)	.41	92ª	0.049 (0.01) ^a	3.7×10^{-7}	88	0.049 (0.051)	.33	94	0.018 (0.050)	.71

Abbreviations: GSMR, generalized summary data-based mendelian randomization; MR, mendelian randomization; NTR, Netherlands Twin Register; QIMR, QIMR Berghofer Medical Research Institute; QSKIN, QSkin Sun and Health

Study; SNPs, single-nucleotide polymorphisms; UKB, United Kingdom Biobank.

place of residence across the United Kingdom, although this association was not replicated in the 2 Australian samples.

Our MR results in the UKB suggest that schizophrenia risk could be a causal factor in the choice to live in more densely populated and low socioeconomic status areas, although more powered analyses would be required to confirm this result in the other cohorts (Figure 3). These results are consistent with the selective migration hypothesis that individuals with genetic liability for schizophrenia tend to move to or remain in urban areas. ³⁸ Larger GWAS for the environmental variables are required to confirm a reverse causation (eTable 9 and eTable 10 in the Supplement) and clarify how psychopathologic traits and residential location relate to each other. More data are needed to clarify the influence of comorbid psychiatric risks ³⁹⁻⁴¹ and associated traits (eg, educational attainment, creativity, or risk taking) ^{11,40,41} on the reported association.

Our work builds on previous research that reported that the density of population of where one lives is significantly heritable⁵ and on nonmolecular studies showing evidence of a familial effect (ie, owing to genetics and/or family environment) in the association between schizophrenia and urban dwelling. ^{4,42,43}

Our results complement 2 recent publications on the interplay between schizophrenia risk and neighborhood. 10,38 The first, from the Swedish registries (N = 759 536), reported an association between schizophrenia PRS and low socioeconomic status neighborhood, 38 which we replicated in the UKB. In addition, we found that SES could not completely explain the association between schizophrenia PRS and population density.

We found a large environmental correlation between SES and population density in Australia (bivariate model including additive genetic, common environmental, and unique environmental factors, $r_{\rm C}$ = 0.86, P < .0001; $r_{\rm E}$ = 0.33, P < .0001) compared with the estimated genetic correlation ($r_{\rm G}$ = 0.35, P = .34.) (eTable 4 in the Supplement). Thus, SES is a potential confounder of genetic analyses of population

density, and conversely, population density is a confounder of SES genetic analyses. Composition of SES measures²³ studied here differed in each country and also differ from the Swedish study.³⁸

A study from the Danish registries found an association between schizophrenia PRS and urban living only for individuals aged 15 years, but not at birth, 10 and did not study the neighborhood during adult age. Herein, we showed that the population density of where a person lives is mostly explained by shared and unique environment, with the heritability increasing with age (eAppendix 11 in the Supplement). Thus, we are uncertain whether shared environment confounded the results observed in the Danish population. However, we cannot rule out a genetic association between upbringing environment and the disease risk (ie, a passive gene-environment correlation) in which the association is driven by the genotype that a child inherits and the environment in which they are raised. Herein, we rather focused on an active gene-environment correlation, presumably driven by selective migration, by including only older participants who have a higher degree of independence in choosing where they live. More work is needed to confirm and examine these results over age groups, which will likely require large, longitudinal cohorts, such as national registries.

We highlighted the importance of age in our analysis by replicating and expanding previous results⁵: that place of residence is heritable and the heritability increases over time, while the influence of family environment declines (eAppendix 11 in the Supplement). This age effect, together with sex differences in prevalence and age of onset of schizophrenia, ^{44,45} justified the study of interactions between PRS and age and sex that may contribute to the choice of neighborhood; however, these interactions were not significant in our analyses.

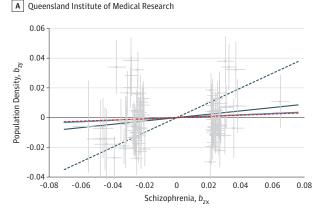
Limitations

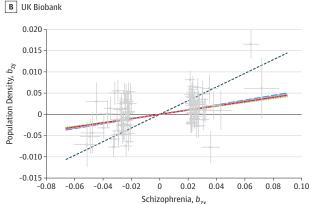
A limitation of our study arises from the low power of the GWAS to detect all variants associated with schizophrenia. 46 This lack

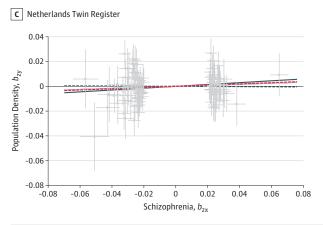
^a Statistically significant results.

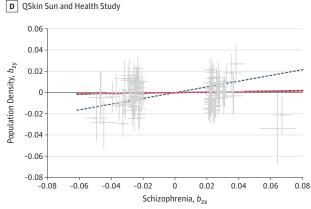
Figure 3. Mendelian Randomization Summary Results











Hypothesis that schizophrenia influences the population density of the place or residence tested in the Queensland Institute of Medical Research (A), UK Biobank (B), Netherlands Twin Register (C), and QSkin Sun and Health Study (D)

cohorts. Several methods yield exactly the same effect sizes (Table 2), and as a consequence, some lines may overlap. GSMR indicates generalized summary data-based mendelian randomization.

of power results in a limited PRS instrument that explains only 11.6% of the total trait heritability in the population. Thus, the variance explained by the current PRS, which is based on common variants, may be only one-tenth of what one would observe with a PRS capturing the whole genetic signal. As a consequence, the small association with population density reported herein may account for 1.7% (95% CI, 0.8%-3.2%) of the schizophrenia risk (based on population density explaining 0.2% of schizophrenia PRS in the QIMR data and 0.002 of 0.116 = 0.017). Ancestry may also confound PRS analyses, ⁴⁷ especially for the variables studied herein (eAppendix 15 and eFigures 17-19 in the Supplement ⁴⁸). We tried to overcome this issue using mixed models that are equivalent to fitting all genetic principal components as covariates (eAppendixes 8 and 15 in the Supplement).

Another limitation is the possible sample overlap between the UKB sample and data used in the schizophrenia GWAS, which may inflate results from PRS and MR analyses (eAppendix 16 in the Supplement). However, we estimated the overlap to be negligible (64 participants or 0.01%, eAppendix 16 in the Supplement), and we did not observe larger effect sizes in the UKB compared with the 3 other cohorts.

Conclusions

Our findings support the notion that the increased schizophrenia prevalence in urbanized areas is not only owing to the environmental stressors of the city or other putative risk factors associated with urbanicity (eg, increased risk of infection, low vitamin D levels, and substance abuse)⁴⁹ but also on the genetic risk for the disease. The associated PRS prediction was replicated across 3 different countries that likely differ in availability of space, social mobility/ opportunities, associations between population density and SES of the area, and historical constraints on living environ-

ment. We showed that the distribution of the genetic risk for the disorder is not uniform and concentrates in more populated and urban areas, supporting the idea of an active gene-environment correlation because of selective migration. Previous evidence of an environmental association between city living and schizophrenia risk^{1,2} is compatible with our results and reflects that there are genetic as well as environmental risk factors for schizophrenia. Furthermore, we provide evidence that schizophrenia genetic risk may lead to individuals (or had led to their ancestors) seeking denser/ urban and low socioeconomic status neighborhoods, which could in turn be risk factors for the disease. ^{1,2,49}

Future disease models will need to include both genetic selection and environmental factors of urban stress on schizophrenia to inform implications for intervention. In addition, there is a need to address the potential gene-by-environment interactions that would arise if genetic variants influencing schizophrenia also influence the choice of a stressful neighborhood, which would contribute to the interaction between urbanicity and family history of schizophrenia that has been reported in the Danish population. ⁵⁰ Such diathesis-stress interaction studies using PRS have been published for depression, ^{51,52} but given the lower prevalence of schizophrenia, national registries will likely be required for investigation.

ARTICLE INFORMATION

Accepted for Publication: May 4, 2018. Published Online: June 23, 2018. doi:10.1001/jamapsychiatry.2018.1581

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Obtained funding: MacGregor, Boomsma, Martin.

Administrative, technical, or material support:

Whitfield, Kemper, Das, MacGregor, Whiteman,

Yang, Martin.

Supervision: Whitfield, Boomsma, Rietschel, McGrath, Medland, Martin.

Conflict of Interest Disclosures: None reported.

Funding/Support: QIMR Phenotype collection, DNA collection, and genotyping were funded by National Health and Medical Research Council (NHMRC) grant 981351 and National Institutes of Health grants RO1 AAO13326, RO1 AAO07535, RO1 AA010249, R01 AA013321, and R37 AA007728 (Dr Martin) over the past 3 decades. This research has been conducted using the UKB Resource under application number 12505. QSKIN is funded by NHMRC project grant APP1063061 (Drs Neale, MacGregor, Olsen, and Whiteman), and NHMRC program grant APP1073898 (Dr Whiteman). Drs Yang, Trzaskowski, Kemper, Yengo, and Couvy-Duchesne were supported by NHMRC grants 1087889 and 1113400. Dr Yang was supported by a Senior Medical Research Fellowship from the Sylvia & Charles Viertel Charitable Foundation. Dr Colodro-Conde was supported by a QIMR Berghofer Fellowship. Dr Couvy-Duchesne was supported by a UQ International scholarship from the University of Queensland (UQ). Dr McGrath was supported by NHMRC John Cade Fellowship APP1056929 and a Niels Bohr Professorship from the Danish National Research Foundation. Dr Neale was supported by NHMRC research fellowship APP1060183. Dr Whiteman was supported by NHMRC research fellowship (APP1058522). Dr Medland was supported by NHMRC research fellowship 1103623.

Role of the Funder/Sponsor: The NHMRC and the Sylvia & Charles Viertel Charitable Foundation did not take part in design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication. QIMR Berghofer and UQ took part in collection and management of the data, but did not take part in the design and conduct of the study; analysis and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Meeting Presentations: Results of the study were presented at the 39th Annual Society for Mental Health Research Conference; December 6, 2017; Canberra, Australia. This paper was presented at the 48th Behavior Genetics Annual Meeting; June 23, 2018; Boston, Massachusetts.

Additional Contributions: The Netherlands
Twin Register acknowledges the Open Data
Infrastructure for Social Science and Economic
Innovations (http://www.odissei-data.nl). We are
grateful to the QIMR, QSKIN, UKB, and NTR
participants, data collectors, and data managers.
Lea Zillich, BSc (University of Heidelberg), provided
comments on the manuscript. She did not receive
financial compensation for the service.

REFERENCES

- 1. Lederbogen F, Kirsch P, Haddad L, et al. City living and urban upbringing affect neural social stress processing in humans. *Nature*. 2011;474 (7352):498-501. doi:10.1038/nature10190
- 2. Lederbogen F, Haddad L, Meyer-Lindenberg A. Urban social stress—risk factor for mental disorders: the case of schizophrenia. *Environ Pollut*. 2013;183: 2-6. doi:10.1016/j.envpol.2013.05.046
- 3. Krabbendam L, van Os J. Schizophrenia and urbanicity: a major environmental influence—conditional on genetic risk. *Schizophr Bull*. 2005;31(4):795-799. doi:10.1093/schbul/sbi060
- 4. Mortensen PB, Pedersen CB, Westergaard T, et al. Effects of family history and place and season of birth on the risk of schizophrenia. *N Engl J Med*. 1999;340(8):603-608. doi:10.1056 /NEJM199902253400803
- 5. Whitfield JB, Zhu G, Heath AC, Martin NG. Choice of residential location: chance, family influences, or genes? *Twin Res Hum Genet*. 2005;8 (1):22-26. doi:10.1375/twin.8.1.22
- **6.** Pedersen CB, Mortensen PB. Evidence of a dose-response relationship between urbanicity during upbringing and schizophrenia risk. *Arch Gen Psychiatry*. 2001;58(11):1039-1046. doi:10.1001/archpsyc.58.11.1039
- **7**. Sundquist K, Frank G, Sundquist J. Urbanisation and incidence of psychosis and depression: follow-up study of 4.4 million women and men in Sweden. *Br J Psychiatry*. 2004;184:293-298. doi:10.1192/bjp.184.4.293
- **8**. Pedersen CB, Mortensen PB. Family history, place and season of birth as risk factors for schizophrenia in Denmark: a replication and reanalysis. *Br J Psychiatry*. 2001;179:46-52. doi:10.1192/bjp.179.1.46
- 9. Marcelis M, Takei N, van Os J. Urbanization and risk for schizophrenia: does the effect operate before or around the time of illness onset? *Psychol Med*. 1999;29(5):1197-1203. doi:10.1017/S0033291799008983

- 10. Paksarian D, Trabjerg BB, Merikangas KR, et al. The role of genetic liability in the association of urbanicity at birth and during upbringing with schizophrenia in Denmark. [published online June 29, 2017]. *Psychol Med*. 2018;48(2):305-314. doi:10.1017/S00332917/17001696
- 11. Ripke S, Neale BM, Corvin A, et al; Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophrenia-associated genetic loci. *Nature*. 2014;511(7510):421-427. doi:10.1038/nature13595
- 12. Wray NR, Lee SH, Mehta D, Vinkhuyzen AA, Dudbridge F, Middeldorp CM. Research review: polygenic methods and their application to psychiatric traits. *J Child Psychol Psychiatry*. 2014; 55(10):1068-1087. doi:10.1111/jcpp.12295
- **13.** Evans DM, Davey Smith G. Mendelian randomization: new applications in the coming age of hypothesis-free causality. *Annu Rev Genomics Hum Genet*. 2015;16:327-350. doi:10.1146/annurev-genom-090314-050016
- 14. Katikireddi SV, Cezard G, Bhopal RS, et al. Assessment of health care, hospital admissions, and mortality by ethnicity: population-based cohort study of health-system performance in Scotland. *Lancet Public Health*. 2018;3(5):e226-e236. doi:10.1016/52468-2667(18)30068-9
- **15.** Heath AC, Bucholz KK, Madden PA, et al. Genetic and environmental contributions to alcohol dependence risk in a national twin sample: consistency of findings in women and men. *Psychol Med.* 1997;27(6):1381-1396. doi:10.1017 /S0033291797005643
- **16.** Knopik VS, Heath AC, Madden PA, et al. Genetic effects on alcohol dependence risk: re-evaluating the importance of psychiatric and other heritable risk factors. *Psychol Med*. 2004;34(8):1519-1530. doi:10.1017/S0033291704002922
- 17. Sudlow C, Gallacher J, Allen N, et al. UK Biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. *PLoS Med*. 2015;12(3):e1001779. doi:10.1371/journal.pmed.1001779
- 18. Bycroft C, Freeman C, Petkova D, et al. Genome-wide genetic data on -500,000 UK Biobank participants [published online July 20, 2017]. bioRxiv. doi:10.1101/166298
- 19. Willemsen G, Vink JM, Abdellaoui A, et al. The Adult Netherlands Twin Register: twenty-five years of survey and biological data collection. *Twin Res Hum Genet*. 2013;16(1):271-281. doi:10.1017/thg.2012.140
- **20**. Olsen CM, Green AC, Neale RE, et al; QSkin Study. Cohort profile: the QSkin Sun and Health Study. *Int J Epidemiol*. 2012;41(4):929-929i. doi:10.1093/ije/dys107
- **21.** Abdellaoui A, Hottenga JJ, de Knijff P, et al. Population structure, migration, and diversifying selection in the Netherlands. *Eur J Hum Genet*. 2013;21(11):1277-1285. doi:10.1038/ejhg.2013.48
- **22.** Whitcher B, Schmid V, Thornton A. Working with the DICOM and NIfTI data standards in R. *J Stat Softw.* 2011;44(6):1-28. doi:10.18637/jss.v044.i06
- 23. Australian Bureau of Statistics. Measures of Socioeconomic Status: ABS catalogue no. 1244.0.55.001. Commonwealth of Australia. http://www.ausstats.abs.gov.au/Ausstats/subscriber.nsf

- /0/367D3800605DB064CA2578B60013445C /\$File/1244055001_2011.pdf. Published 2011. Accessed May 24, 2018.
- **24**. Boker S, Neale M, Maes H, et al. OpenMx: an open source extended structural equation modeling framework. *Psychometrika*. 2011;76(2): 306-317. doi:10.1007/s11336-010-9200-6
- **25.** *R: A Language and Environment for Statistical Computing.* Vienna, Austria: R Foundation for Statistical Computing; 2012.
- **26**. Neale MC, Cardon LR. *Methodology for Genetic Studies of Twins and Families*. Kluwer Academic Publishers; 1992. doi:10.1007/978-94-015-8018-2
- 27. Purcell S. Variance components models for gene-environment interaction in twin analysis. *Twin Res.* 2002;5(6):554-571. doi:10.1375/136905202762342026
- **28**. Purcell S, Neale B, Todd-Brown K, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*. 2007;81(3):559-575. doi:10.1086/519795
- **29.** Loh PR, Tucker G, Bulik-Sullivan BK, et al. Efficient Bayesian mixed-model analysis increases association power in large cohorts. *Nat Genet*. 2015;47(3):284-290. doi:10.1038/ng.3190
- **30**. Loh P-R, Kichaev G, Gazal S, Schoech AP, Price AL. Mixed model association for biobank-scale data sets [published online September 27, 2017]. *bioRxiv*. doi:10.1101/194944
- **31.** Feng S, Liu D, Zhan X, Wing MK, Abecasis GR. RAREMETAL: fast and powerful meta-analysis for rare variants. *Bioinformatics*. 2014;30(19):2828-2829. doi:10.1093/bioinformatics/btu367
- **32.** Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet*. 2011;88(1):76-82. doi:10.1016/j.ajhg.2010.11.011
- **33.** Yang J, Zaitlen NA, Goddard ME, Visscher PM, Price AL. Advantages and pitfalls in the application of mixed-model association methods. *Nat Genet*. 2014;46(2):100-106. doi:10.1038/ng.2876
- **34.** Bulik-Sullivan BK, Loh PR, Finucane HK, et al; Schizophrenia Working Group of the Psychiatric Genomics Consortium. LD score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet*. 2015; 47(3):291-295. doi:10.1038/ng.3211
- **35**. Hemani G, Zheng J, Wade KH, et al. MR-Base: a platform for systematic causal inference across the phenome using billions of genetic associations [published online December 16, 2016]. *bioRxiv*. doi: 10.1101/078972
- **36**. Hemani G, Haycock P, Zheng J. TwoSampleMR: Two Sample MR functions and interface to MR Base database. https://mrcieu.github.io/TwoSampleMR/. Published 2017. Accessed May 24, 2018.
- **37**. Zhu Z, Zhang Z, Zhang F, et al. Causal associations between risk factors and common diseases inferred from GWAS summary data. *Nat Commun*. 2018;9(1): 224. doi:10.1038/s41467-017-02317-2
- **38.** Sariaslan A, Fazel S, D'Onofrio BM, et al. Schizophrenia and subsequent neighborhood deprivation: revisiting the social drift hypothesis using population, twin and molecular genetic data. *Transl Psychiatry*. 2016;6:e796. doi:10.1038/tp.2016.62

- **39**. Bulik-Sullivan B, Finucane HK, Anttila V, et al; ReproGen Consortium; Psychiatric Genomics Consortium; Genetic Consortium for Anorexia Nervosa of the Wellcome Trust Case Control Consortium 3. An atlas of genetic correlations across human diseases and traits. *Nat Genet*. 2015; 47(11):1236-1241. doi:10.1038/ng.3406
- **40**. Anttila V, Bulik-Sullivan B, Finucane HK, et al. Analysis of shared heritability in common disorders of the brain [published online September 6, 2017]. *bioRxiv*. doi:10.1101/048991
- **41**. Power RA, Steinberg S, Bjornsdottir G, et al. Polygenic risk scores for schizophrenia and bipolar disorder predict creativity. *Nat Neurosci.* 2015;18(7): 953-955. doi:10.1038/nn.4040
- **42**. Cantor-Graae E, Selten JP. Schizophrenia and migration: a meta-analysis and review. *Am J Psychiatry*. 2005;162(1):12-24. doi:10.1176/appi.ajp.162.1.12
- **43**. Pedersen CB, Mortensen PB. Are the cause(s) responsible for urban-rural differences in schizophrenia risk rooted in families or in individuals? *Am J Epidemiol*. 2006;163(11):971-978. doi:10.1093/aje/kwj169
- **44.** Abel KM, Drake R, Goldstein JM. Sex differences in schizophrenia. *Int Rev Psychiatry*. 2010;22(5):417-428. doi:10.3109/09540261.2010 .515205
- **45**. McGrath J, Saha S, Welham J, El Saadi O, MacCauley C, Chant D. A systematic review of the incidence of schizophrenia: the distribution of rates and the influence of sex, urbanicity, migrant status and methodology. *BMC Med*. 2004;2:13. doi:10.1186/1741-7015-2-13
- **46**. Dudbridge F. Power and predictive accuracy of polygenic risk scores. *PLoS Genet*. 2013;9(3): e1003348. doi:10.1371/journal.pgen.1003348
- **47**. Curtis D. Polygenic risk score for schizophrenia is more strongly associated with ancestry than with schizophrenia [published online March 23, 2018]. *bioRxiv*. doi:10.1101/287136
- **48**. Haworth S, Mitchell R, Corbin L, et al. Common genetic variants and health outcomes appear geographically structured in the UK Biobank sample: old concerns returning and their implications [published online April 11, 2018]. *bioRxiv*. doi:10.1101/294876
- **49**. Plana-Ripoll O, Bocker Pedersen C, McGrath JJ. Urbanicity and risk of schizophrenia—new studies and old hypotheses [published online May 16, 2018]. *JAMA Psychiatry*. doi:10.1001/jamapsychiatry.2018.0551
- **50**. van Os J, Pedersen CB, Mortensen PB. Confirmation of synergy between urbanicity and familial liability in the causation of psychosis. *Am J Psychiatry*. 2004;161(12):2312-2314. doi:10.1176/appi.ajp.161.12.2312
- **51.** Mullins N, Power RA, Fisher HL, et al. Polygenic interactions with environmental adversity in the aetiology of major depressive disorder. *Psychol Med.* 2016;46(4):759-770. doi:10.1017/S0033291715002172
- **52**. Colodro-Conde L, Couvy-Duchesne B, Zhu G, et al; Major Depressive Disorder Working Group of the Psychiatric Genomics Consortium. A direct test of the diathesis-stress model for depression. [published online July 11, 2017]. *Mol Psychiatry*. 2017. doi:10.1038/mp.2017.130